

## Fungi isolated from the rhizosphere of spring cruciferous plants

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Fungal communities isolated from the rhizosphere of spring cruciferous plants were analysed in the study. It was found that the rhizosphere of crucifers was colonized primarily by fungi of the order *Mucorales* and of the genus *Fusarium*. Members of the genus *Fusarium* dominated in the rhizoplane. The roots of cruciferous plants secrete glucosinolates – secondary metabolites known for their antifungal properties, thus affecting the communities of soil-dwelling fungi.

**Key words:** rhizosphere, cruciferous plants, fungi, *Fusarium*

### INTRODUCTION

The soil provides habitat for both phytopathogenic and saprotrophic microorganisms, including bacteria, actinomycetes and fungi (Patkowska 1998). The rhizosphere, i.e. the zone that surrounds the roots of plants, plays a particularly important role due to its specific biological properties. It is teeming with a wide variety of microbes (Morgan et al. 2005), which can be divided into plant growth-promoting rhizobacteria (PGPR), deleterious rhizosphere microorganisms (DRMO) and neutral microorganisms – having no impact on plant growth (Sturz, Christie 2003). Under natural conditions, in undisturbed soil, the groups of beneficial and harmful microorganisms remain in the state of dynamic equilibrium. PGPR contribute to yield increment, usually resulting from higher nutrient availability and suppression of the growth and activity of deleterious microorganisms. DRMO compete with PGPR for food, thus negatively affecting crop development (Kurek, Kobus 1990). Due to their antagonist potential, soil microorganisms are able to colonize suitable niches. The antagonistic mechanisms include antibiosis, competition and mycoparasitism (hyperparasitism) (Sturz, Christie 2003). Many pathogens develop in the after-harvest residues of forecrops, so the type of forecrop may have a significant influence on the

yield of successive crops (Bojarczuk, Bojarczuk 1988). Crucifers and legumes play a positive role in crop rotation because they improve the chemical, physical and biological properties of the soil (Majchrzak et al. 2002). Members of the family *Brassicaceae* are among the best forecrops, because they leave in the soil large amounts of after-harvest residues rich in glucosinolates and other secondary metabolites (Oleszek 1997). Moreover, they exert a positive effect on the health of successive crops (Majchrzak et al. 2004; Majchrzak et al. 2005).

The objective of this study was to determine the species composition of fungi isolated from the rhizosphere of selected cruciferous plants.

## MATERIALS AND METHODS

The study was conducted during the years 1999-2001 at the Production-Experimental Station in Bałcyny near Ostróda (NE Poland), on the experimental plots of the Department of Plant Production, University of Warmia and Mazury in Olsztyn. The experiment was established on gray-brown podsolic soil developed from light silty clay, of quality class III a, of good wheat complex (1999 and 2000) or very good rye complex (2001). Crucifers were grown after spring wheat in 1999 and 2000, and after winter rapeseed in 2001. Mineral fertilizers (NPK) were applied at the following rates: 60-100 : 40-60 : 60-100 kg•ha<sup>-1</sup>, as recommended by the Institute of Soil Science and Plant Cultivation. The field trial was performed in a randomized block design, in three replications. The experimental factors were as follows:

- factor I – cruciferous plants
- spring oilseed rape (*Brassica napus f. annua*) – cv. Margo,
  - white mustard (*Sinapis alba*) – cv. Heter,
  - Chinese mustard (*Brassica juncea* var. *sareptana*) – cv. Małopolska,
  - radish (*Raphanus sativus* var. *oleifera*) – cv. Pegletta,
  - false flax (*Camelina sativa*) – cv. Borowska,
  - Spanish colewort (*Crambe abyssinica*) - cv. Borowski
- factor II – years of the study

No fungicides were applied. Fungi were isolated from the rhizosphere, rhizoplane and roots of crops as described by Mańska (1974). The quantitative and qualitative composition of fungal communities was determined at full blooming (BBCH 65-69).

## RESULTS

A total of 2 929 fungal colonies belonging to 99 species and non-spore forming fungi were isolated from the rhizosphere of spring cruciferous plants over the three-year experimental period (Tab.1). Members of the order *Mucorales* dominated among them (48.96% of all colonies). Representatives of the genus *Rhizopus* were isolated most frequently (15.94%). Fungi of the genus *Penicillium* were also abundant (12.43%). Antagonistic species, including the order *Mucorales* and the genera *Gliocladium*, *Penicillium* and *Trichoderma*, accounted for 66.58% of all isolates. The proportion of pathogens in the rhizosphere was 11.33%, and the predominant role was played by species of the genus *Fusarium* (8.77%), primarily *F. solani*, *F. oxysporum* and *F. equiseti*.

The most diverse fungal community, composed of 637 colonies representing 44 species, was isolated from the rhizosphere of spring rape (Tab. 1). *Mucorales* (46.10%), *Fusarium* (16.5%), *Penicillium* (7.85%) and *Gliocladium* (6.12%) dominated among them. The fewest fungi (339 isolates belonging to 36 species) were detected in the rhizosphere of false flax. The most abundant among them were: *Mucorales* – 57.8%, *Acremonium* – 9.44% and *Fusarium* – 6.49%. Fungi of the genus *Fusarium* were isolated least frequently from the rhizosphere of Spanish colewort (3.81%). The genus *Penicillium* (270 isolates – 46.80%) dominated in the rhizosphere of this crop. The rhizospheres of white mustard, Spanish colewort and radish were characterized by the lowest species diversity (35 fungal species each).

A total of 568 fungal colonies belonging to 80 species and non-spore forming fungi were isolated from the rhizoplane of spring cruciferous plants during the experimental period (Tab. 2). The most diverse fungal community, comprising 134 colonies representing 34 species, was isolated from the rhizosphere of spring rape. Members of the genera *Fusarium* (32.84%) and *Acremonium* (21.64%) as well as of the order *Mucorales* (5.69%) dominated among them. The fewest fungi were isolated from the rhizoplane of Spanish colewort (73 isolates). This community, composed of only 24 species, was found to be the least diverse. The rhizoplane of this crop was mostly colonized by *Gliocladium* spp. (21.90%), *Aspergillus fumigatus* (15.10%) and *Penicillium* spp. (15.10%). Representatives of the genus *Fusarium* constituted the least numerous group in the rhizoplane of Spanish colewort (6 isolates – 8.22%).

## DISCUSSION

Research results show that plants of the genus *Brassicaceae* grown as forecrops or ploughed in as green manure have a beneficial effect on the health of field crops (Majtahedi et al. 1991). The roots of crucifers secrete glucosinolates, which affect the soil microflora and help to control the occurrence of phytopathogens (Bones, Rossiter 1996; Kierkegaard, Sarwar 1998). Decomposition of the tissues of *Brassicaceae* as well as the production of glucosinolates followed by their hydrolysis lead to the formation of isothiocyanates (ITCs) – volatile substances considered to be biofumigants (Sarwar et al. 1998). According to Snapp et al. (2007) and Charron and Sam (1999), growing plants of the genus *Brassicaceae* as forecrops and leaving their remainders in the field inhibits the growth of such soil pathogens as *Rhizoctonia solani* and *Pythium ultimum*. Marwar and Lodha (2002) demonstrated that plants of the family *Brassicaceae* limited the occurrence of *Fusarium oxysporum* f. sp *cumini*.

In the present study fungi of the genus *Fusarium* were not abundant in the rhizosphere of crucifers. Their population was considerably greater in the rhizoplane. The soil environment of particular cruciferous plants was colonized by members of this genus to a different degree. *Fusarium* colonies were isolated most frequently from the rhizosphere and rhizoplane of spring oilseed rape, and least frequently from the rhizosphere and rhizoplane of Spanish colewort and false flax. Ishimoto et al. (2000) reported that fungi of the genus *Fusarium* isolated from the roots of crucifers showed high tolerance to glucosinolates, which may suggest that they acquired resistance to this group of substances through adaptation.

In the current experiment the rhizosphere of the *Cruciferae* was colonized by numerous representatives of the order *Mucorales*, dominated by members of the

Table 1  
Fungi isolated from rhizosphere spring cruciferous plants

Fungal species	Plants										Sum	%
	Oilseed rape	White mustard	Chinese mustard	Oilseed rape	False flax	Spanish colewort	1	2	3	1		
<i>Acremonium breve</i> (Sukap. et Thirum) W. Gams	1*	2	3	1	2	3	1	2	3	1	2	3
<i>Acremonium charticola</i> (Lindau) W. Gams	21	33	2	3	4	3				11	6	
<i>Acremonium chrysogenum</i> (Sukap.&Thirum) W. Gams										2		2
<i>Acremonium curvulum</i> W. Gams	16		6						6			28
<i>Acremonium kiliense</i> Gruetz							15		8			23
<i>Acremonium larvarum</i> (Petch) W. Gams							14					14
<i>Acremonium minutisporum</i> (Sukap. et Thirum) W. Gams								6				6
<i>Acremonium ochraceum</i> (Onions et Barron) W. Gams								1				1
<i>Acremonium porroni</i> Vuill.	1									4		5
<i>Acremonium psammosporum</i> W. Gams	4			2			2		11			19
<i>Acremonium sclerotigenum</i> (F. et R. Moreau ex Valenta) W. Gams			1									1
<i>Acremonium strictum</i> W. Gams	1										3	3
<i>Acremonium</i> sp.	1		4		1	27	5	3		1	42	143
<i>Alternaria alternata</i> (Fr.) Keissler					1	3				4		4
<i>Arthrinium sphaerospermum</i> Fuckel						1						1
<i>Aspergillus repens</i> de Bary					2	2	1					1
<i>Aureobasidium bolleyi</i> Sprague								7	7			5
<i>Aureobasidium pullulans</i> de Bary				14		1					3	31
<i>Botyodiplodia</i> sp.											1	1
<i>Borytis cinerea</i> Persoon	3		5			1	1		4		8	27
<i>Cephalosporium atrum</i> de Bary												5
<i>Cercospora</i> spp.											5	0.17
<i>Cladosporium cladosporoides</i> (Fres.) de Vries	18	10	6	2	8	10	15	5			74	253
<i>Cladosporium herbarum</i> Link ex FR.		2		1			11	2				16
<i>Colletotrichum</i> spp.							5					5
<i>Coniothyrium</i> spp.	1		5	3			4					13
<i>Cylindrocampus destructans</i> (Zins.) Scholten		4	6	2	2	1					15	0.51
<i>Endothia</i> sp.		3									3	0.10
<i>Epicoccum purpurascens</i> Ehrenb. ex Schlecht						1					1	0.03
<i>Epiconium</i> spp.											1	0.03
<i>Fusarium avenaceum</i> Corda ex Fr.	1		3				2		1		16	0.55
<i>Fusarium elatamydosporum</i> (Wollenweber et Reinking)		2						2			4	0.14
<i>Fusarium culmorum</i> (W.G. Smith) Sacc.									1		2	0.07

Fungal species	Plants												Sum %					
	Oilseed rape			White mustard			Chinese mustard			Oilseed rape			False flax			Spanish colowort		
	1*	2	3	1	2	3	1	2	3	1	2	3	1	2	3	1	2	3
<i>Fusarium dimerum</i> (Penzig)																		
<i>Fusarium equiseti</i> (Corda) Sacc.	5	10	4	3	1					12	5		2	1		3	46	1.57
<i>Fusarium nivale</i> (Fr.) Ces.		1								13	1		1			16	0.55	
<i>Fusarium oxyoporum</i> (Schlecht.)	13	29	2	7	2	8	1			12	4	1	4	9	2	94	3.21	
<i>Fusarium semibucinum</i> Fuck.																		
<i>Fusarium solani</i> (Mart.) Sacc.	18	1	2	17	1	2	1			4	7		1			3	3	0.10
<i>Fusarium solani</i> var. <i>coryneum</i> Thüm																4	58	1.98
<i>Fusarium tabacinum</i> (Beyma) W. Gams																2	0.07	
<i>Fusarium</i> sp.																		
<i>Gliocladium catenulatum</i> Gilman et Abbott	1						2			10	2				11		26	0.89
<i>Gliocladium fimbriatum</i> Gilman et Abbott		4													5		9	0.31
<i>Gliocladium penicilloides</i> Corda	11	22	12	7						2	1				4		59	2.01
<i>Gliocladium salmoneicolor</i> Raillo				1												1	0.03	
<i>Gliomastix cerealis</i> (Karl.) Dickinson	1						1										1	0.03
<i>Gliomastix murorum</i> (Corda) Hughes								3									1	0.03
<i>Humicola fusco-atra</i> Traaen	5			2													6	0.96
<i>Humicola brevis</i> Gilman et Abbott				5												5	14	0.48
<i>Humicola nigrescens</i> Omvik				1	2			3			1	1				8	0.27	
<i>Klebsiella alttistrina</i> Coemans																1	0.03	
<i>Monocillium mucidum</i> W. Gams																2	2	0.07
<i>Monodictis levis</i> (Wilsh.) Hughes		2	2				2									6	0.20	
<i>Mortierella calliacea</i> Linnemann			1													1	0.03	
<i>Mortierella elongata</i> Linnemann													2			2	0.07	
<i>Mortierella gemmifera</i> Ellis																1	0.03	
<i>Mortierella gracilis</i> Linnemann	7															7	0.24	
<i>Mortierella marburgensis</i> Linnemann				2											2	4	0.14	
<i>Mortierella alpina</i> Peyroni	13			7												30	1.02	
<i>Mortierella vinacea</i> Dixon -Stewart																1	1	0.03
<i>Mortierella</i> spp.																		
<i>Mucor circinelloides</i> van Tieghem																2	2	0.07
<i>Mucor hiemalis</i> Wehmeyer	86	36	7	72	14	2	253			11	115	11	15	5		30	18	675
<i>Mucor microsporus</i> Namyslowskii					6											6	0.20	
<i>Mucor mucredo</i> (Linne) Brefeld				3												2	5	0.34
<i>Mucor piriformis</i> Fischer																37	1.26	
<i>Mucor pusillus</i> Lindt										23			5				28	0.96
<i>Mucor racemosus</i> Fresenius	8			7						5		11				31	1.06	
<i>Paecillomyces variabilis</i> Barron												1				1	0.03	

Table 1  
Fungi isolated from rhizosphere spring cruciferous plants – cont.

Fungal species	Plants										Sum %									
	Oilseed rape			White mustard			Chinese mustard			Oilseed rape			False flax			Spanish colewort				
	1*	2	3	1	2	3	1	2	3	1	2	3	1	2	3	1	2	3		
<i>Penicillium nigricans</i> (Bain.) Thom																			0.10	
<i>Penicillium</i> spp.	35	15		12	5	1	16			2	1	4	1	230	39	361	12.33			
<i>Periconia funerea</i> (Ces.) Mason et Ellis		9					2									2	0.07			
<i>Phoma europaea</i> Sacc.										6						15	0.51			
<i>Phoma finetii</i> (Brun)							9		2							11	0.38			
<i>Phoma glomerata</i> (Corda) Wollenweber et Hochapel	2															2	0.07			
<i>Phoma herbarum</i> Westend.		8														2	0.07			
<i>Rhizoctonia solani</i> Kühn								1		1						8	0.27			
<i>Rhizoctonia</i> spp.																2	0.07			
<i>Rhizopus nigricans</i> Ehrenberg																9	0.31			
<i>Rhizopus</i> spp.	113			71			39		39	5	152					39	458	15.64		
<i>Sclerotinia sclerotiorum</i> (Lib.) de Bary							15													
<i>Scopulariopsis acremonium</i> (Delacr.) Vuill.	2							1								15	0.51			
<i>Scopulariopsis brevicaulis</i> (Sacc.) Bain								13								3	0.10			
<i>Scyphularidium lignicola</i> Pesantie	4															13	0.44			
<i>Spicaria carnosaa</i> Miller; Giddens et Foster									1							3	7	0.24		
<i>Spicaria divaricata</i> (Thun.) Gilman&Abbott							7			9						7	0.24			
<i>Spicaria griseoolla</i> Saccardo								1								9	0.31			
<i>Spicaria simplicissima</i> Oudemans									32							1	0.03			
<i>Spicaria violacea</i> Abbott	41			1				1								73	2.49			
<i>Sporotrichum carnis</i> Brooks et Hansford					1											2	0.07			
<i>Sporotrichum chlorinum</i> Link										2						1	0.03			
<i>Sporotrichum olivaceum</i> Fries										5						2	0.07			
<i>Torula herbarum</i> (Pers.) Link ex Fr.	10								5							5	0.17			
<i>Trichoderma aureoviride</i> Rifai																2	0.58			
<i>Trichoderma hamatum</i> (Bon.) Bain	6		2	5	1		10			1					1	16	0.55			
<i>Trichoderma harzianum</i> Rifai		1													8	19	0.65			
<i>Trichoderma polysporum</i> (Link et Pers.) Rifai		5													5	0.17				
<i>Verticillium cellulosae</i> Daszewska			3												3	0.10				
Non-sporulating fungi	1	1	4	4				5	1	5	5				32	1.09				
Sum	98	299	240	106	203	107	29	339	190	87	189	126	57	88	194	67	368	142	2929	100.0
Sum of plant	637	416						558		402					577					

\*1 - 1999 2 - 2000 3 - 2001

Table 2  
Fungi isolated from rhizoplane of spring cruciferous plants

Fungal species	Plants												Sum %							
	Oilseed rape			White mustard			Chinese mustard			Oilseed rape			False flax			Spanish colewort				
	1*	2	3	1	2	3	1	2	3	1	2	3	1	2	3	1	2	3		
<i>Acremonium breve</i> (Sukap.&Thirum.) W. Gams	8									2	2	1						8	1.41	
<i>Acremonium charicola</i> (Lindau) W. Gams	2						1												8	1.41
<i>Acremonium chrysogenum</i> (Sukap. et Thirum.) W. Gams	1			1															2	0.35
<i>Acremonium curvulum</i> W. Gams				7															7	1.23
<i>Acremonium incoloratum</i> (Sukap. et Thirum.) W. Gams	1																		1	0.18
<i>Acremonium kilense</i> Gruetz	1			1															3	
<i>Acremonium larvarum</i> (Petch) W. Gams																			5	0.88
<i>Acremonium minutisporum</i> (Sukap. et Thirum.) W. Gams								1										1	0.18	
<i>Acremonium poroni</i> Vuill.	2			1														1	2	0.35
<i>Acremonium psammosporum</i> W. Gams	1			1														5	0.88	
<i>Acremonium strictum</i> W. Gams	1			1														2		0.35
<i>Acremonium</i> spp.	3			1														9	1.58	
<i>Acrospeira mirabilis</i> Berk. et Br.																		2	0.35	
<i>Alternaria alternata</i> (Frits.) Keissler				1			1			9	1		2		2			17	2.99	
<i>Aspergillus fumigatus</i> Fresenius								1										11	2.11	
<i>Aspergillus funiculosus</i> (G. Smith)																		1	0.18	
<i>Aspergillus repens</i> de Bary																		1	0.18	
<i>Ascochyta</i> spp.				1			2											3	0.53	
<i>Aureobasidium bolleii</i> (Sprague)				1			1			1		1						1	0.18	
<i>Aureobasidium pullulans</i> de Bary				1			6	1	3	1		1		2	2		1	19	3.35	
<i>Botryotrichum pululiferum</i> Saccardo et Marchal				1														1	0.18	
<i>Botryos cinerea</i> Persoon							1											3	0.53	
<i>Cladosporium cladosporoides</i> (Fres.) de Vries	1			2	1	1				4	4	1			6	1		22	3.87	
<i>Cladosporium herbarum</i> (Persoon) Link					1			1							3			6	1.06	
<i>Coniothyrium</i> spp.					1			1										2	0.35	
<i>Cylindrocarpon destructans</i> (Zins.) Scholten				2	1		1		1	1							1	7	1.23	
<i>Endothia</i> sp.				1														1	0.18	
<i>Epicoccum</i> sp.				1														1	0.18	
<i>Fusarium avenaceum</i> (Corda ex Fr.)	1			1														4	0.70	
<i>Fusarium culmorum</i> (W.G. Smith) Sacc.							2			1							1	5	0.88	
<i>Fusarium equisetii</i> (Corda) Sacc.				1				1			1						1	5	0.88	
	6	1					3				1	3					15	2.64		

Table 2  
Fungi isolated from rhizoplane of spring cruciferous plants

Fungal species	Plants										Sum %							
	Oilseed rape			White mustard			Chinese mustard			Oilseed rape			False flax			Spanish colewort		
	1*	2	3	1	2	3	1	2	3	1	2	3	1	2	3	1	2	3
<i>Fusarium fusariooides</i> (Frag. Cif.)																		
<i>Fusarium oxysporum</i> (Schlecht.)	3	1	1	1	1	1	3	2	3	1	2	3	1	2	3	1	2	3
<i>Fusarium solani</i> (Mart.) Sacc.	1	15	1	4	3	4	2	2	2	2	2	3	5	3	5	2	21	370
<i>Fusarium solani</i> var. <i>coeruleum</i> Thüm	1						7			1		2	3	2		1	40	704
<i>Fusarium tabacinum</i> (Beyma) W. Gams et Abbott	1																9	158
<i>Gliocladium catenulatum</i> Gilman et Abbott	8									1	1	1	1	1	1	1	2	0.35
<i>Gliocladium fimbriatum</i> Gilman & Abbott							1									14	2.46	
<i>Gliocladium penicilloides</i> Corda		2	1				2			1						13	14	2.46
<i>Gliomastix murorum</i> (Corda) Hughes		1		1												6	1.06	
<i>Humicola fuscoc-aura</i> Traenæ	1	4	2			2			3	1		2					1	0.18
<i>Humicola brevis</i> Gilman et Abbott					1				1							16	2.82	
<i>Humicola nigrescens</i> Omvik																2	0.35	
<i>Kickxella alabastrina</i> Coemans						1											3	0.53
<i>Microdochium niveale</i> (Fr.) Ces.				11						1		2					1	0.18
<i>Monocillium arcticola</i> (W. Gams)									1								14	2.46
<i>Monierella alpina</i> Peyron	1	1											1				1	0.18
<i>Monierella arcuata</i> Wolf												1					1	0.18
<i>Monierella isabelina</i> Oudemans	1			1													1	0.18
<i>Monierella vinacea</i> Dixon -Stewart																	1	0.18
<i>Mucor circinelloides</i> van Tieghem																	11	1.94
<i>Mucor hiemalis</i> Wöhmer																	3	0.53
<i>Mucor mucero</i> (Linne) Brefeld																	52	9.15
<i>Mucor pififormis</i> Fischer																	3	0.53
<i>Mucor racemosus</i> Fresenius																	2	0.35
<i>Paecilomyces variotii</i> Bainier																	4	0.70
<i>Paecilomyces niveus</i> Stolk et Samson	3	2	7				11		30	2	2	1	3	1	1		1	0.18
<i>Penicillium nigricans</i> (Bain.) Thom										2							1	0.18
<i>Penicillium</i> spp.		2	8	7			2		2		7	4	1	4	1	8	2	8.45
<i>Phoma chrysanthemica</i> Hollos														1	1		1	0.18
<i>Phoma europea</i> Sacc.										2	2		1				5	0.88
<i>Phoma glomerata</i> (Corda) Wollenweber et Hochapel											2						2	0.35
<i>Phoma herbarum</i> Westend.																1	1	0.18

Fungal species	Plants										Sum %	
	Oilseed rape	White mustard	Chinese mustard	Oilseed rape	False flax	Spanish colowort						
1*	2	3	1	2	3	1	2	3	1	2	3	
<i>Pyrenophora semeniperda</i> (Brittlebank et Adam)												
<i>Rhizoctonia solani</i> Kühn	1			1							0.70	
<i>Rhizoctonia</i> spp.	2										0.18	
<i>Rhizophus</i> spp.	1				3	15	6	5	4		0.35	
<i>Scopulariopsis brevicaulis</i> (Sacc.) Bain							1			2	6.34	
<i>Scopulariopsis brumpii</i> Salvanet-Duval										1	0.18	
<i>Scyathidium lignicola</i> Pesante						1				1	0.18	
<i>Spicaria elegans</i> Corda			3							1	0.18	
<i>Spicaria simplicissima</i> Oudemans	1		1							1	0.18	
<i>Spicaria violacea</i> Abbott		3		6	2					2	0.35	
<i>Sporotrichum carnis</i> Brooks et Hansford		1								13	2.29	
<i>Tonella herbarum</i> (Pers.) Link ex Fr.						1				1	0.18	
<i>Trichoderma aureoviride</i> Rifai				1	1				1	1	0.18	
<i>Trichoderma hamatum</i> (Bon.) Bain			1	1	3	2				3	0.53	
<i>Trichoderma harzianum</i> Rifai						1					7	1.23
<i>Trichoderma polysporum</i> (Link et Pers.) Rifai			1							1	0.18	
Non-sporulating fungi										1	0.18	
spring cruciferous plants	24	78	32	22	26	7	80	46	11	6	16	
Sum of plant	134	74	74	74	133	79	79	75	73	28	568 100.0	

\*1 - 1999 2 - 2000 3 - 2001

genus *Rhizopus*. Ishimoto et al. (2000) confirmed the predominant role of the genus *Rhizopus* in the rhizosphere of cruciferous plants. According to these authors, fungi of the genus *Rhizopus* showed significantly higher tolerance for glucosinolates than fungi of the genus *Fusarium*.

## CONCLUSIONS

1. The largest and the most diverse fungal community was isolated from the soil environment of spring oilseed rape.
2. The fungal populations that colonized the rhizosphere and rhizoplane of Spanish colewort and false flax were found to be the smallest.
3. Members of the order *Mucorales* dominated in the soil environment of cruciferous plants.
4. Fungi of the genus *Fusarium* were isolated least frequently from the soil environment of Spanish colewort and most frequently from the soil environment of spring oilseed rape.

## REFERENCES

- Bojarczuk M., Bojarczuk J. 1988. Fitosanitarna ocena wartości przedplonów roślin zbożowych. *Fragmenty Agronomii*. 1(17): 5–23.
- Bones A.M., Rossiter J.T. 1996. The myrosinase-glucosinolate system, its organisation and biochemistry. *Physiol. Plant.* 97:194–208.
- Charron C.S., Sams C.E. 1999. Inhibition of *Pythium ultimum* and *Rhizoctonia solani* by shredded leaves of *Brassica* species. *J. Am. Soc. Hortic. Sci.* 124:462–467.
- Ishimoto H., Fukushi Y., Yoshida T., Tahara S. 2000. *Rhizopus* and *Fusarium* are selected as dominant fungal genera in rhizospheres of *Brassicaceae*. *Journal of Chemical Ecology* 26 (10): 2387–2399.
- Kirkegaard J.A., Sarwar M. 1998. Biofumigation potential of brassicas. I. Variation in glucosinolate profiles of diverse field-grown brassicas. *Plant Soil* 201:71–89.
- Kurek E., Kobus J. 1990. Korzystne i szkodliwe oddziaływanie mikroflory ryzosferowej na wzrost i rozwój roślin. *Postępy Mikrobiologii* 29 (1/2): 103–123.
- Majchrzak B., Kurowski T.P., Karpińska Z. 2002. Zdrowotność jarych roślin krzyżowych a grzyby zasiedlające ich nasiona. *Acta Agrobot.* 55 (1): 199–210.
- Majchrzak B., Okorski A., Chodorowski B. 2004. Zdrowotność korzeni i podstawy źdźbła pszenicy jarej uprawianej po różnych przedplonach. *Annales UMCS* 59(4): 1779–1788.
- Majchrzak B., Chodorowski B., Okorski A. 2005. Choroby podstawy źdźbła pszenicy ozimej uprawianej po roślinach przedplonowych z rodziny *Brassicaceae*. *Acta Agrobot.* 59 (2): 307–318.
- Majtahedi H., Santo G.S., Hang A.N., Wilson J.H. 1991. Suppression of root-knot nematode populations with selected rapeseed cultivars as green manure. *J. Nematol.* 23:170–174.
- Mańska K.. 1974. Zbiorowiska grzybów jako kryterium oceny wpływu środowiska na choroby roślin. *Zeszyty Prob. Post. Nauk Roln.* 160: 9–22.
- Marwar R., Lodha S. 2002. *Brassica amendments* and summer irrigation for the control of *Macrophomina phaseolina* and *Fusarium oxysporum* f. sp *cumini* in hot arid region. *Phytopathol. Mediterr.* 41:45–54.
- Morgan J. A. W., Bending G. D., White P. J. 2005. Biological costs and benefits to plant-microbe interactions in the rhizosphere. *Journal of Experimental Botany* 56 (417): 1729–1739.
- Oleszek W. 1997. Glukozynolany - występowanie i znaczenie ekologiczne. *Wiadomości Bot.* 39 (1/2): 49–58.
- Patkowska E. 1998. Wpływ resztek roślinnych na zdrowotność i plonowanie soi. *Roczn. Akad. Roln. Poznań, Ogrod.* 27: 213–219.
- Sarwar M., Kirkegaard J.A., Wong P.T.W., Desmarchelier J.M. 1998. Biofumigation potential of Brassicas. III. In vitro toxicity of isothiocyanates to soil-borne fungal pathogens. *Plant Soil* 201:103–112.

- Snapp S.S., Date K.U., Kirk W., O'Neil K., Kremen A., Bird G. 2007. Root, shoot tissues of *Brassica juncea* and *Cereal secale* promote potato health. *Plant Soil.* 294:55–72.
- Sturz A. V., Christie B. R. 2003. Beneficial microbial allelopathies in the root zone: the management of soil quality and plant disease with rhizobacteria. *Soil & Tillage Research.* 72: 107–123.

### Grzyby ryzosferowe jarych roślin kapustnych

#### Streszczenie

W badaniach poddano analizie zbiorowiska grzybów ryzosferowych jarych roślin kapustnych. Ryzosferę roślin kapustnych zasiedlały przede wszystkim grzyby z rzędu *Mucorales* i rodzaju *Fusarium*. W ryzoplanie roślin znacznie częściej występowali przedstawiciele rodzaju *Fusarium*. Korzenie rośliny kapustnych wydzielają do gleby glukozynolany, które wpływają na zbiorowiska grzybów zasiedlających środowisko glebowe roślin.