

Stenocarpella macrospora AND *Stenocarpella maydis* IN THE CERRADO AND SOUTHERN BRAZIL REGIONS

Stenocarpella macrospora E *Stenocarpella maydis* NAS REGIÕES DO CERRADO E SUL DO BRASIL

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ABSTRACT: *Stenocarpella macrospora* and *Stenocarpella maydis* may result in the seedlings death or cause rotting at the corn stalk base and in all or part of the ear. In addition, *S. macrospora* can cause leaf spot. Double-haploid strains from corn hybrids resistant to *S. macrospora* and *S. maydis* were identified. Also the incidence of these pathogens in the Cerrado and in Southern Brazil localities was determined. One hundred and forty double-haploid maize hybrids, in addition to the controls, were inoculated with *S. macrospora* and *S. maydis* and evaluated for resistance reaction in three locations of the Cerrado and three locations of the South regions. The grains attacked by these fungi were collected and variable quantities of *S. macrospora*, *S. maydis* and other fungal species were registered. The results demonstrated the prevalence of *S. macrospora* in the Cerrado as well as other non-*Stenocarpella* sp. fungi in the South. The city of Abelardo Luz (Santa Catarina) was the only place where *S. maydis* was found to have a higher incidence than *S. macrospora*. Environmental effects influence the prevalence of fungi, causing grain rot. These results indicated genetic gains in the selection of hybrids resistant to this fungi for use as direct breeders in *Stenocarpella*-corn pathological system research.

KEYWORDS: *Diplodia* sp. Rotten grain. Double-haploids.

INTRODUCTION

Corn (*Zea mays* L.) is second in economic importance and cultivated area in the world. *Stenocarpella maydis* (Berk.) Sacc., *Stenocarpella macrospora* Earle, *Fusarium verticillioides* Sheld and *Fusarium graminearum* (Schw) are the principal agents of disease in terms of rot in the stalk and ear. These fungi reduce crop yield and depreciate product quality due to the production of toxins (EDDINS, 1930; PROZESKY et al., 1994; DORRANCE et al, 1998;. ROSSOUW et al, 2002a;. HOUSE, et al, 2006;. GUTIERREZ, 2008).

In the natural environment, *S. macrospora* and *S. maydis* occur only in imperfect or asexual form (HOUSE et al., 2006). On summer the climatic conditions of Southern Brazil with warm days (25-27°C) and mild nights (12-15°C) are favorable *S. maydis* development (PEREIRA, 1995). In certain environments where the relative humidity is less than 50% *S. macrospora* produces more mycelium and pycnidia, growing faster than *S. maydis* and can infect plants at any phenological stage (DEL RIO, 1990).

In the ear symptoms usually begin shortly after fertilization. When infection occurs two weeks after pollination, the ear can become completely affected with a brownish-gray to off-white color in the fungus. Infected grains have a dull gray to black color and brown pycnidia can form on the tassel,

floral bracts, ears and grains (HOUSE et al, 2006;. WOLOSSHUK; WISE, 2008).

Differentiation between these species can require breeding programs to use specific germplasm for each pathogen, according the program scope. However, there were few studies that focus on tropical germplasm and fewer that aim to identify resistance of these two pathogens (MARIO; REIS; JULIATTI, 2011). Thus, the present study aimed to identify resistance to *S. macrospora* and *S. maydis* of double-haploid corn hybrids and also verify the incidence of these pathogens in the Cerrado and in Southern Brazil regions. Results would provide valuable information to breeding programs on studies of the *Stenocarpella*-corn pathosystem.

MATERIAL AND METHODS

Hybrids, environments and experimental design

The hybrids examined resulted from a cross between two contrasting inbred lines conducted to determine their reaction to rot ear by *Stenocarpella* spp., MLR1 (resistant) and MLS1 (susceptible), both produced by from Monsanto.

One hundred and forty double-haploid lines were generated using a haploid frequency inductor in combination with colchicine treatment. These lines were crossed with a susceptible line tester (MLS4) and were unrelated to parental hybrids in

order to generate an equal number of test intersections.

For chromosomal replication, seeds were immersed in a .06% colchicine solution and .5% dimethylsulfoxide (DMSO) for 12 h in the dark (DEIMLING, 1997) at room temperature. After duplication, the seedlings were washed for 20 min in running water and taken to the greenhouse.

The resulting hybrids were evaluated during the 2006/07 season in the Cerrado and Southern Brazil regions, in a totalizing six locations. In the Cerrado, evaluations were conducted in Irai de Minas, Uberlândia and Araguari, in the State of Minas Gerais. In the South, the trials were conducted in Pinhão and Guarapuava, in Paraná, and at Abelardo Luz, in Santa Catarina. Selection of these environments was mainly based on the historical incidence of grain rot obtained from each of the tested environment research programs. The adoption of direct sowing on straw for consecutive cycles was examined as a common practice as well as representative yield levels of the sampled regions.

The experimental design included randomized blocks with two replications per location. The experimental unit consisted of two rows of five m, spaced 70 cm between each row. The plant density was 70,000 and 75,000 plants ha⁻¹ in the Cerrado region and in the South, respectively. Mechanical sowing and harvesting were adopted. Plants were fertilized with 185-80-100 kg ha⁻¹ (N-P-K) in two doses. The first dose was applied at sowing, using 50-80-100 kg ha⁻¹ (N-P-K). The second dose, of 135 kg ha⁻¹ of nitrogen, was applied 30 days after sowing. For weed control three liters per hectare of a mixture of atrazine (200g L⁻¹) and metalachlor (200g L⁻¹) were applied.

Origin of the isolates and insulation

The isolates used for the artificial inoculation were collected from infected ears in the region of Uberlândia. Isolation of the *S. maydis* and *S. macrospora* was conducted in the Plant Pathology Laboratory (Monsanto Company, Uberlândia, MG).

Grains with typical symptoms of the disease were placed in a humid chamber for seven days at 25°C and 95% relative humidity to stimulate pycnidia formation. Using a stereoscopic microscope and histological needle, pycnidia were collected from the grain, placed on a drop of water and covered with a slide. The conidia were examined for species identification and transferred to a Petri dish containing potato-dextrose-agar culture medium (PDA). The fungal species were

incubated for three days at 23-27°C. Subsequently, the resulting colonies were transferred to new Petri dishes containing PDA and incubated for 3-4 days at 25°C.

Inoculation

One hundred grams of grain sorghum were washed in plain water using a one L Erlenmeyer flask. The washed grains were placed in 125 mL of distilled water for 12 h; the unabsorbed water was discarded. The substrate was then autoclaved twice at 125°C for 20 min. Five discs of *Stenocarpella* spp. with 5,0 mm diameter were transferred to the sorghum substrate. The flasks were then incubated at 25°C until spore masses formed around the sorghum grains. They were then kept in a shaker for five days for equal distribution.

The inoculum was suspended in 250 mL of distilled water, agitated for 30 min and transferred to another flask through a funnel containing 5,0 layers of cheesecloth for filtration. The conidia were counted in a Newbauer chamber and the suspension was adjusted to a concentration of 4x10⁴ conidia mL⁻¹ (MÁRIO; REIS; JULIATTI, 2011).

Inoculation was performed throughout the experimental plot using the inoculum method which consisted of sprinkling a suspension of spores of *S. macrospora* and *S. maydis* over the floral bracts on the ear peduncles with the aid of an automatic dosing syringe. Each ear received 5,0 mL of suspension containing 4x10⁴ conidia mL⁻¹ of each specie. The inoculation was performed 10 to 15 days after the plants had reached 100% of female flowering (MÁRIO, 1998; SMITH et al, 2005).

Sample collection and data analysis

The harvest was carried out when the grain reached 18-24% of moisture. Each plot produced a grain sample of approximately 300 g to estimate the percentage of rotted grain and identify the pathogens by the filter paper method using one hundred grains per repetition. The hybrid grain yields (kg ha⁻¹) were also calculated.

Five resistant, 5 moderately resistant and 5 susceptible hybrids from each region of the study were evaluated for *S. macrospora* and/or *S. maydis* incidence. The control genotypes, CheckR and CheckS, were also evaluated. Any fungi collected from rotted grain, that were not *Stenocarpella* spp. were classified as "other fungi", beyond the focus of the present study. Hybrids were evaluated in three locations of the Cerrado (Savana conditions) and in three locations from southern Brazil. Two joint analyses were conducted, one in the three locations of the Cerrado region and another in three locations

of the South, thus, six repetitions were analyzed by location.

The detection of *S. macrospora* and *S. maydis* in the grain was performed based on the color differences of the colonies, using filter paper method (MÁRIO; REIS, 2001) and growing at the growth chamber set at 25 °C and 12 h photoperiod.

Analysis of Variance

After the incubation period (chamber set at 25 °C) the percentage of rotted grains per observation in two replications and randomized blocks was evaluated. After this was determined the percentage of grains with the both *Stenocarpella* species. The distribution and characterization of the incidence of rotted grain in the hybrids studied were evaluated by descriptive statistics and histograms. To estimate averages of the rotted grain incidence and their genetic components, we adopted the analysis model (SAS Institute Inc, vs 9.2), with completely randomized blocks:

$$y_{ij} = \mu + b_j + t_i + e_{ij}$$

Where: y_{ij} is the value observed in the j repetition from the test-cross I ; μ is a constant inherent in all observations; b_j is the random effect of the j location; $b_j \sim N(0, \sigma_b^2)$, t_i is the test-cross i t_i

$\sim N(0, \sigma_g^2)$ effect and e_{ij} is the random error associated with y_{ij} , $e_{ij} \sim N(0, \sigma_e^2)$ observation.

Components of genetic, residual and phenotypical variances were estimated by the moments method, by the solution of equations obtained comparing the mathematical expectations of the means squared means of the analysis of variance, to their observed values, determined by:

Genetic variance component:

$$\hat{\sigma}_g^2 = \frac{GenotypeMS - ResidueMS}{r};$$

Residual variance component: $\hat{\sigma}^2 = ResidueMS$;

Phenotypic variance component: $\hat{\sigma}_f^2 = \hat{\sigma}_g^2 + \frac{\hat{\sigma}^2}{r}$;

Heritability: $\hat{h}^2 = \frac{\hat{\sigma}_g^2}{\hat{\sigma}_f^2}$.

RESULTS AND DISCUSSION

Joint analysis of the 140 double-haploid hybrids tested in the two regions of Brazil showed significant statistical differences for the test crossing (hybrid) effect (Table 1). This indicated the existence of significant positive variance between hybrids derived from MLR1 / MLS1 crossing in the Cerrado region and in Southern Brazil (Table 1).

Table 1. Joint analysis of variance for incidences of rotted grain in double-haploid corn hybrids tested in Cerrado and Southern Brazil regions.

Evaluated Regions	Residual Variance	Genetic Variance	G x L Variance	Phenotypic Variance	Heritability
Cerrado	9.40	1.21	.81	3.05	.46
Southern Brazil	7.26	0.96	.62	2.37	.43

*G x L: genotype x location

The joint analysis results for each region (Table 2) allowed classification of the hybrids according to the incidence of rotted grain. Overall, there was variation in the resistance of the hybrids by region showing an environmental effect on the expression of the trait.

The joint analysis of the Cerrado (Table 2) produced higher percentages of rotted grain incidence (MH55, MH48, MH89, MH77 and

MH128) ranging from 9.52 to 13.16%. This was a higher incidence than found in the hybrid used as a rotted grain susceptibility control (CheckS) (7.34%). The most resistant hybrids for the Cerrado were: MH2, MH9, MH134, MH106 and MH41, with a rotted grain incidence ranging from 3.06% to 3.36%. The hybrid control used as resistance reference (CheckR) was the most resistant (2.89%).

Table 2. Joint analysis by region. Incidences of rotted grain and corn grain yield data.

Hybrids	Cerrado		Hybrids	Southern Brazil	
	Rotten grain (%)	Grain Yield (kg ha ⁻¹)		Rotten grain (%)	Grain Yield (kg ha ⁻¹)
MH1	5.89	9.104	MH1	9.03	10.858

MH2*	3.06	8.599	MH2*	4.06	11.693
MH3	4.40	7.598	MH3	5.09	11.958
MH4	6.62	8.345	MH4	6.51	10.885
MH5**	6.35	8.701	MH5	6.89	11.932
MH6	6.86	8.010	MH6	6.41	9.009
MH7	7.31	7.521	MH7	8.03	8.662
MH8	6.00	9.234	MH8	4.86	11.665
MH9*	3.17	9.222	MH9	5.18	11.195
MH10	6.61	9.696	MH10	7.64	11.320
MH11	5.57	8.456	MH11	6.59	9.632
MH12	5.24	8.857	MH12	6.12	10.740
MH13	5.67	7.943	MH13	6.97	10.506
MH14	4.65	8.328	MH14	5.76	10.487
MH15	7.41	8.524	MH15	8.27	9.477
MH16	7.42	8.553	MH16	8.11	10.947
MH17	7.30	8.777	MH17	6.89	10.386
MH18	7.81	7.777	MH18	7.46	11.312
MH19	7.59	8.678	MH19	7.32	11.988
MH20	8.22	9.153	MH20	6.06	10.424
MH21	7.79	7.735	MH21	7.31	11.861
MH22	5.73	8.821	MH22	5.81	10.903
MH23	9.22	7.533	MH23	6.21	10.494
MH24	6.67	9.105	MH24	6.58	11.321
MH25	5.48	8.766	MH25	8.17	9.942
MH26	6.95	7.694	MH26	6.17	12.040
MH27	6.60	7.978	MH27	5.49	10.610
MH28	3.88	8.724	MH28	5.35	10.678
MH29	6.10	9.023	MH29	5.44	10.703
MH30	5.07	9.929	MH30	5.04	11.253
MH31	5.00	7.905	MH31	7.43	11.789
MH32	5.55	9.151	MH32	5.79	12.336
MH33	5.84	9.045	MH33	7.29	11.463
MH34	8.75	8.845	MH34	5.78	11.504
MH35**	6.25	8.149	MH35	5.45	11.092
MH36	4.42	7.810	MH36	6.12	9.689
MH37	5.12	7.280	MH37	7.98	10.710
MH38	7.41	8.879	MH38***	10.85	10.940
MH39	5.35	8.820	MH39**	6.69	11.905
MH40	5.47	8.263	MH40	5.97	10.533
MH41*	3.36	8.844	MH41*	4.56	10.493
MH42	4.61	8.757	MH42	5.83	10.934
MH43	7.06	9.229	MH43	8.96	10.668
MH44	4.18	7.890	MH44	5.35	11.643
MH45	6.20	7.154	MH45	5.95	9.482
MH46	3.97	7.412	MH46	4.85	11.662

MH47	6.66	8.105	MH47	7.10	10.640
MH48***	9.58	8.225	MH48	7.20	10.845
MH49	5.31	8.182	MH49	6.03	10.190
MH50	3.82	8.539	MH50	5.58	11.054
MH51	3.87	8.362	MH51	6.59	11.353
MH52	6.21	8.054	MH52*	4.60	11.368
MH53	5.07	8.478	MH53	7.86	10.476
MH54	7.52	7.526	MH54	6.11	9.435
MH55***	9.52	7.213	MH55	5.28	10.781
MH56	6.74	9.098	MH56	8.42	11.707
MH57	6.54	7.733	MH57	5.71	9.763
MH58	4.00	9.250	MH58	5.82	11.308
MH59	7.62	7.738	MH59	7.34	10.358
MH60	7.77	8.531	MH60	7.52	9.467
MH61	5.07	7.853	MH61	4.79	11.276
MH62	4.62	8.545	MH62	6.50	11.094
MH63	4.74	7.116	MH63	9.42	10.794
MH64	5.45	8.655	MH64	5.06	11.115
MH65	5.23	7.104	MH65	6.11	10.795
MH66	7.83	6.992	MH66	7.44	11.447
MH67	8.66	8.316	MH67	7.78	9.148
MH68	6.10	7.646	MH68	5.97	9.967
MH69	7.96	5.239	MH69	8.45	10.104
MH70**	6.26	8.327	MH70	7.68	11.021
MH71	6.13	7.709	MH71	4.91	10.497
MH72	3.88	8.451	MH72	4.85	10.116
MH73	4.66	6.767	MH73	5.17	9.845
MH74	7.76	7.856	MH74	8.43	11.217
MH75	6.14	7.458	MH75	7.40	9.461
MH76	6.23	9.686	MH76	5.62	11.646
MH77***	12.71	7.896	MH77	8.11	8.916
MH78	7.36	6.840	MH78	5.71	9.558
MH79	5.53	7.510	MH79	5.49	11.173
MH80	7.95	7.431	MH80	8.77	10.087
MH81	8.00	8.727	MH81	6.38	10.911
MH82	3.54	8.375	MH82	4.99	11.452
MH83**	6.27	7.638	MH83	8.11	10.306
MH84	8.67	9.488	MH84**	6.64	10.818
MH85	5.67	8.589	MH85	7.66	11.311
MH86	6.75	7.721	MH86	8.87	11.587
MH87	7.06	10.002	MH87**	6.79	11.336
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MH88	6.36	9.428	MH88	5.94	11.827
MH89***	11.50	5.550	MH89***	14.25	8.055
MH90	5.57	7.763	MH90	5.64	10.518
MH91	6.07	8.982	MH91	8.40	11.784

MH92	5.79	8.206	MH92	5.07	11.467
MH93	5.66	8.981	MH93	6.96	11.177
MH94	4.95	8.852	MH94	8.48	11.785
MH95	5.71	8.373	MH95***	11.42	11.313
MH96	8.31	9.005	MH96***	10.16	10.128
MH97	7.33	8.358	MH97	7.22	10.733
MH98	7.10	7.349	MH98	5.29	11.214
MH99	6.45	9.155	MH99	6.99	9.604
MH100	4.11	9.556	MH100	6.51	11.309
MH101	5.46	8.759	MH101	5.62	9.486
MH102	7.73	8.042	MH102	7.21	9.501
MH103**	6.27	7.133	MH103	8.07	10.630
MH104	7.50	9.405	MH104**	6.71	12.205
MH105	4.60	9.221	MH105	8.21	11.889
MH106*	3.29	8.881	MH106	7.15	11.045
MH107	4.52	7.243	MH107	6.48	11.862
MH108	7.12	8.876	MH108	5.35	11.767
MH109	4.61	8.075	MH109	4.81	10.554
MH110	6.73	9.475	MH110	5.73	10.344
MH111	5.24	7.988	MH111**	6.87	9.472
MH112	4.79	8.254	MH112*	4.57	10.722
MH113	6.21	8.358	MH113	4.61	11.214
MH114	6.47	9.170	MH114	6.04	10.973
MH115	4.61	9.764	MH115	5.34	12.642
MH116	7.38	7.389	MH116	6.87	10.090
MH117	8.36	7.534	MH117	7.94	10.805
MH118	7.62	8.430	MH118	8.23	11.022
MH119	9.35	9.576	MH119	6.58	11.680
MH120	6.83	8.705	MH120	9.33	10.640
MH121	5.72	8.655	MH121	4.78	11.122
MH122	6.40	8.684	MH122	5.88	10.394
MH123	5.76	7.516	MH123	8.23	9.868
MH124	5.84	7.856	MH124	5.26	11.689
MH125	5.72	8.228	MH125***	10.74	9.945
MH126	8.31	8.121	MH126	8.50	10.180
MH127	4.40	9.187	MH127	6.51	10.399
MH128***	13.16	7.251	MH128	8.24	9.889
MH129	7.08	8.509	MH129	6.24	11.094
MH130	8.53	8.918	MH130	7.43	11.378
MH131	7.24	8.282	MH131	9.58	12.004
MH132	5.37	9.579	MH132	6.09	11.084
MH133	5.52	9.157	MH133*	4.60	11.782
MH134*	3.22	9.610	MH134	5.91	10.926
MH135	6.00	7.644	MH135	6.51	10.158
MH136	6.02	9.556	MH136	6.01	11.858

MH137	8.49	7.303	MH137	9.40	10.528
MH138	5.31	7.464	MH138	5.80	9.647
MH139	11.61	1.869	MH139	7.36	7.312
MH140	6.04	7.941	MH140	6.87	11.338
CheckR	2.89	10.044	CheckR	4.40	11.848
CheckS	7.34	9.229	CheckS	7.52	11.748

Corn hybrids selected in each region for analysis of the health of damaged kernels: * Five hybrids with high resistance; ** Five hybrids with moderate resistance; *** Five hybrids with low resistance. Hybrids studied: MH 1-140. CheckR: resistance control. CheckS: susceptibility control. Experimental Hybrids from Monsanto Company, Uberlândia, MG.

In the Southern Brazil joint analysis (Table 2), the most susceptible hybrids were: MH96, MH125, MH38, MH95 and MH89. These hybrids presented incidences ranging from 10.16% to 14.25%, a higher incidence than the CheckS (7.52%). Hybrids with lower disease percentages were MH2, MH41, MH112, MH52 and MH133. The MH2 hybrid was the most resistant (4.06%), better than the resistance control CheckR (4.40%).

The MH2 and MH41 hybrids exhibited disease resistance in both regions of the experiment. On the other hand, MH89 was found to be susceptible in both regions (Figure 2). The results indicated stability of the characteristics for these hybrids regardless region or climate where they were tested.

The hybrids expressed better productive potential when grown in the southern region (Table

2). Cerrado region production ranged from 1,869 to 10,044 kg ha⁻¹ while in the South it ranged from 7,312 to 12,642 kg ha⁻¹.

The Figure 1 represents incidences of *S. macrospora*, *S. maydis* and other fungi in the selected hybrids from the analysis of each region. The fungus *S. maydis* had the lowest incidence in all treatments, ranging from 10 to 13%. Regardless region and environment, a prevalence of *S. macrospora* compared to *S. maydis*, was found in the rotted grain samples. The fungus *S. macrospora* had higher incidences when hybrids were tested in the Cerrado, with a grain rot incidence of 60%, compared to 58% of grain rot incidence in the South. On the other hand, the percentage of other fungi, that also cause ear rot, was higher in the South (69%) than in the Cerrado (61%).

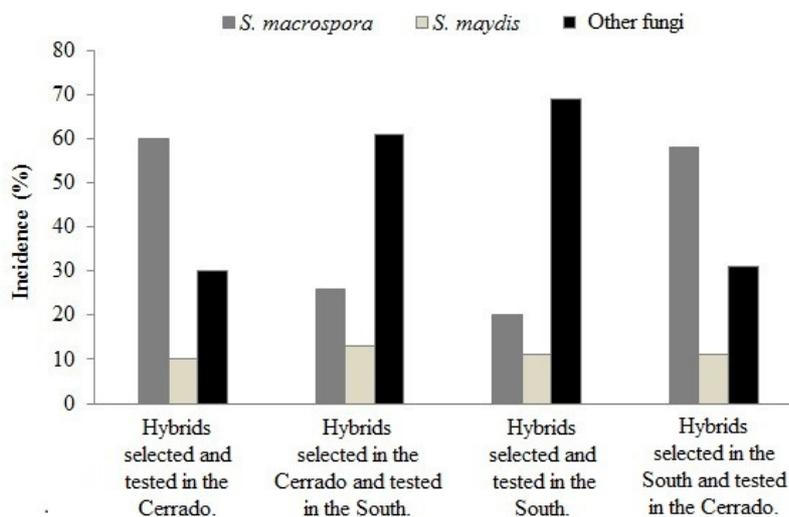


Figure 1. Incidence of rotted grain caused by *S. macrospora*, *S. maydis* and other fungi in hybrids selected in a joint analysis of the Cerrado and Southern Brazil regions.

Between the three Cerrado localities (Figure 2), Iraí de Minas presented the highest incidence of *S. macrospora* (72%) but also a lower *S. maydis* incidence (3%). In Uberlândia 52% of the samples contained *S. macrospora*, 13%, *S. maydis* and 35% other fungi that cause rotting in grain; Araguari registered 43%, 23% and 34%, respectively.

In the South the three locations studied included Abelardo Luz, Pinhão and Guarapuava.

Abelardo Luz exhibited a higher incidence of *S. maydis* (39%) than *S. macrospora* (17%) (Figure 3). Pinhão and Guarapuava had low *S. maydis* percentages, 1 and 2%, respectively, compared to *S. macrospora*: 31 and 15%, respectively. Thus, it was observed that Abelardo Luz was the only location where *S. maydis* presented a higher incidence than of *S. macrospora* (Figures 2 and 3).

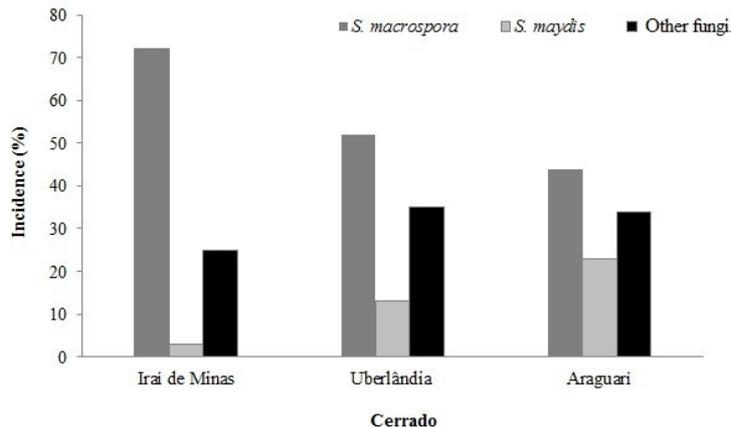


Figure 2. Incidence of rotted grain caused by *S. macrospora*, *S. maydis* and other fungi in hybrids selected from three localities in the Cerrado Region.

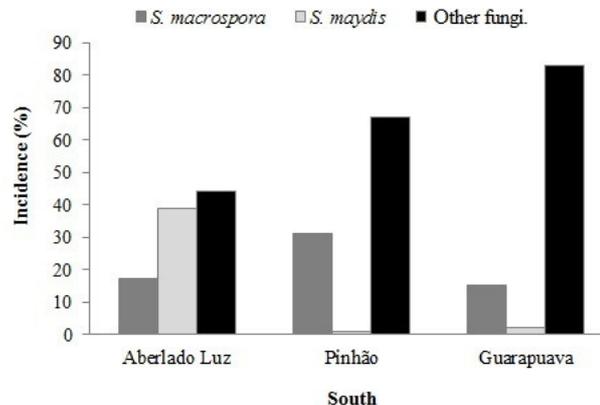


Figure 3. Incidence of rotted grain caused by *S. macrospora*, *S. maydis* and other fungi in hybrids selected from three localities of Southern Brazil region.

In summary, five resistant hybrids, five of medium strength and five susceptible hybrids were

selected for running the joint analysis for each region (Table 2) and were used to verify incidences

of *S. macrospora*, *S. maydis* and other fungi (Figures 1 to 3). The hybrid MH139, when analyzed in the Cerrado region, showed higher susceptibility than the hybrids MH55, MH48 and MH89 that had been selected for analysis but MH139 had low productivity (Table 2). Reasons for this result were not determined and will remain for subsequent analyzes.

The *Stenocarpella* spp. inoculation method utilized in this study permitted differentiation of resistant and susceptible corn genotypes. Studies have shown that natural inoculation efficiency is higher when compared to other methods such as the colonized-stick-tooth, that may cause difficulties in the assessment because of injuries caused when the colonized stick is introduced into the corn ear (BENSCH et al., 1992; MARIO et al., 2003; SILVA, et al., 2005).

In the South and Cerrado regions joint analysis, a significantly positive variance was obtained for the tested hybrids. The heritability coefficients were .46 and .43 in the Cerrado and the South, respectively (Table 1). A trend indicating higher disease expression in the Cerrado was thus suggested. Other studies have also verified genotype and environmental interaction, indicating the need to conduct grain rot resistance evaluations in different locations (DORRANCE et al., 1998; ROSSOUW et al., 2002a, 2002b).

The present study with 140 double-haploid corn hybrids demonstrated that it was possible to detect variations in rotted grain incidence (Table 2). Double haploid progenies presented high resistance to *Stenocarpella* spp. when compared to the resistant control (CheckR) and susceptible checking (CheckS). Differential responses from hybrids have been reported in terms of leaf spot and damage caused by *Stenocarpella* spp. Under ideal environmental conditions for disease hybrids classified as resistant may still have some degree of susceptibility (THOMPSON et al., 1971; MARIO; PRESTES, 1997; TRENT et al., 2002; PASCUAL, et al., 2006). These results can be verified, for example, by the resistance control (CheckR), which had varying values between the Cerrado region (2.89%) and the South (4.40%) (Table 2).

The hybrids evaluated showed higher yields in the South (Table 2). However, the yield was not affected by grain quality. Similar studies have indicated that cob rot is also a factor that significantly reduces grain quality rather than grain yield. This is unlike cob rot diseases that can cause yield reduction (THOMPSON et al., 1971; TRENT et al., 2002; MARIO et al., 2003).

Stenocarpella spp. grain rot incidence values showed a continuous distribution (data not presented), indicating the possibility that this resistance is conditioned by more than one factor. These results are consistent with previous studies reporting multiple genetic aspects as responsible for grain rot resistance when the rot is caused by *Stenocarpella* spp. (OLATINWO et al., 1999, GUTIERREZ, 2008).

Two hybrids in particular showed grain rot percentage values similar to the control results, in both regions: MH2 and MH41 (Table 2). This indicated that there may be genetic gains in grain rot selection incited by *Stenocarpella* spp. Certainly these will serve breeding programs as sources of *Stenocarpella* spp. resistance.

When incidences of *S. macrospora*, *S. maydis* and other fungi were tested and evaluated a higher incidence of *S. macrospora*, in relation to *S. maydis* (Figure 1) was observed in both regions.

These results are similar of those obtained by Mario et al., (2003), who found a fourfold incidence of *S. macrospora* (12.36%) compared to *S. maydis* (3.25%). This may be related to the fact that *S. macrospora* can infect corn leaves in addition to the grains and thus, the leaf spot conidia may serve as an inoculum source to increase incidences of rotted grain. Leaves are positioned near the infection site, the corn ear peduncle (MARIO; PRESTES, 1997; MARIO; REIS, 2003). Other research has reported that increases and intensity of stalk and grain rot may be associated with inoculum density, especially in leaves lesions (DEL RIO, 1990; FLETT; MCLAREN, 1994; HOUSE, 2000). Moreover, a higher incidence of *S. macrospora* can be attributed to greater inoculum availability in crop residues, where spores may have been released and carried by the wind to infection sites (MARIO; REIS, 2003).

Although the summer climatic conditions of southern Brazil (warm days and balmy nights) favor *S. maydis* development (PEREIRA, 1995), only in Abelardo Luz was the *S. maydis* incidence (39%) higher than the incidence of *S. macrospora* (17%) (Figure 3). According to Reis and Mario, 2003, most research in Brazil refers to *S. maydis* as one of the most frequent pathogens in corn, but this research have found similar incidences for both species. According to these authors, diagnoses based on mycelium coloring can generate equivocal results and incorrect diagnosis between the two pathogens. However, when using the method described by Mario and Reis (2001), pathogen identification is possible with a greater certainty, in order to recognize the differences between these two

Stenocarpella species. Furthermore, it should be noted that in similar climatic situations, Brazil and South Africa for example, one of the main fungi associated with rotted grain in corn crop is *Stenocarpella macrospora* (MARASAS; VAN DER WESTHUIZEN, 1979).

CONCLUSIONS

There was a significant difference in the mean incidence of rotted grain between the Cerrado (savana conditions) and southern Brazil regions.

This suggested that there are genetic gains for this variable in hybrid selection and breeding programs.

The results identified environment effects on the prevalence of fungi that cause rotting in corn. There was a prevalence of *S. macrospora* in the Cerrado and other fungi non-*Stenocarpella* ssp. in southern Brazil.

Abelardo Luz was the only location where there was a higher incidence of *S. maydis* than of *S. macrospora*.

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RESUMO: *Stenocarpella macrospora* e *Stenocarpella maydis* em milho, podem resultar na morte de plântulas ou causar apodrecimento na base do caule e da totalidade ou parte da espiga. Além disso, *S. macrospora* pode causar manchas foliares. Identificou-se linhagens duplo-haplóides de híbridos de milho resistentes a *S. macrospora* e *S. maydis*; determinou-se também a incidência desses patógenos no Cerrado e do Sul do Brasil. Cento e quarenta híbridos duplo-haplóides de milho além dos controles (testemunhas) foram inoculados com *S. macrospora* e *S. maydis* e avaliados quanto à resistência em três localidades do Cerrado e três de Sul do Brasil. Os grãos atacados pelos fungos foram colhidos e avaliados quanto à incidência dos dois patógenos. Foram estimadas as porcentagens (%) de *S. Macrospora* e de *S. Maydis* e também a ocorrência de outros fungos pelo método de *blotter*. Houve maior presença de *S. macrospora* do Cerrado. No Sul do Brasil, o município de Abelardo Luz foi o único local onde *S. maydis* foi encontrado em maior incidência do que *S. macrospora*. Os resultados mostraram efeitos ambientais sobre a prevalência de fungos que causam grãos ardidos. Estes resultados indicaram ganhos genéticos na seleção de híbridos resistentes ao fungo *S. Macrospora* e obtenção de híbridos resistentes em milho, tanto na região do Cerrado como no Sul do Brasil.

PALAVRAS-CHAVE: *Diplodia* sp. Grãos ardidos. Duplo-haplóides

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