

MOBILITY AND DISTRIBUTION OF WHEY SOLUTIONS IN SOIL COLUMNS

MOBILIDADE E DISTRIBUIÇÃO DE SOLUTOS DE SORO DE LEITE EM COLUNAS DE SOLO

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ABSTRACT: In Brazil's food industry, dairy production is one of the most important sectors, whose most relevant byproduct is whey. Due to the difficulties of reuse and environmental impacts caused when discarded as effluent in water bodies, an alternative for its final destination would be the application of this residue in the soil. The purpose of this study was to determine chemical changes and mobility and distribution of solutes in the soil after applications of whey rates, as well as to analyze the leachate collected after each application. The test was carried out in a laboratory, in PVC columns filled with soil. The treatments consisted of 2 x 2 samples of a typical dystrophic Red-Yellow Oxisol (Oxisol) and a typical dystrophic fluvic Inceptisol (Inceptisol), sampled in the layers 0- 20 and 20-40 cm. Each experimental unit consisted of 11 PVC rings (diameter of 6.6 cm, height of 7 cm). The columns were arranged in a randomized complete block design with five replications. Four whey rates were applied, corresponding to a soil pore volume of 0.2, at intervals of six days. The leachate was collected 24 and 120 hours after each application to measure pH, electrical conductivity (EC), chemical oxygen demand (COD), contents of total N, N-NH₄⁺, N-NO₃⁻, Na, K, Ca, and Mg. Six days after the last whey application, the columns were opened and the soil of each ring was analyzed for pH, EC, total N, N-NH₄⁺, N-NO₃⁻, Na, K, Ca, and Mg. The high electrolyte concentrations of whey resulted in a general increase in soil EC. The increase of N-NH₄⁺ and N-NO₃⁻ in the soil was high due to mineralization. High concentrations of K, Na and Ca caused displacement of Mg from the exchange complex. It was concluded that from an environmental standpoint, whey soil application is a viable alternative, given that problems of salinization and leaching of undesirable elements are avoided by an adequate management.

KEYWORDS: Nitrogen mineralization. Leaching. Salinization.

INTRODUCTION

Whey consists of a milk co-product that is extracted from milk coagulation in the cheesemaking process (BRAZIL, 2013). This residue stands out due to the large volume generated (about 9 L kg⁻¹ of cheese manufactured) and its quality characteristics, which allow several possibilities of use.

Whey represents approximately 80 to 90% of the milk volume and contains 55% of its nutrients, the water content varies from 91 to 95% (SILVA, 2006). Dry extract averages 6.9%, of which 5% corresponds to lactose; 0.9% to proteins; 0.3% to lipids; and 0.1% to lactic acid, in addition to other nutrients present in lower concentrations (SANTOS, 2001).

The direct use of whey in human consumption includes its use mainly in the manufacture of dairy drinks and in the production of ricotta. The beneficiation processes allow the wide use of this residue in the manufacture of other products in the food industry and even in the pharmaceutical industry (MAHMOUD, 1994; BOUNOUS et al., 1999). However, the

recovery of whey substances for such purposes depends on the availability of appropriate techniques and the use of economically viable processes.

Studies indicate potential use of whey in pig feed, mainly because of its high palatability and nutritional value. However, experimental studies indicate that whey can substitute up to 20% of the pig diet that are in the growing and finishing stages, without compromising performance and carcass characteristics (BERTOL et al., 1993).

As for cattle feeding, it is recommended that whey should not be substituted for milk at the stage of lactation and that it should be supplied in a limited way to adult animals. Some problems arising from the supply of whey to cattle are common, such as palatability drop after two days of storage due to souring. Diarrhea and tympanism can occur if animals consume large quantities of whey in a short period of time or if they are not consuming an adequate amount of whey (LIZIEIRE; CAMPOS, 2001).

The presence of proteins, lipids, mineral salts and lactose confers high polluting power to the whey

when disposed without criteria in the environment (ROLFS et al., 2011). The treatment of whey as effluent in wastewater treatment plants is generally hampered due to the high daily volume and the high concentration of organic matter, ie high levels of biochemical oxygen demand (BOD), mainly due to the presence of lactose (MACHADO et al., 2001) Thus, most of the time, the destination given to this effluent is the disposal in the waterways.

Whey BOD varies from 30 to 60 g L⁻¹, depending on the specific processing used in cheeses and lactose content (BRANDÃO, 1994). Therefore, the discharge of serum directly into watercourses leads mainly to the consumption of dissolved oxygen by microorganisms, and the mortality of fish and other individuals in the aquatic ecosystem may occur (ARAÚJO, 2001).

Given the above, studies become necessary to enable new forms of whey utilization. The disposition in the soil appears as an alternative when it is not possible to use this by-product in human or animal food.

In countries like the United States, the world leader in milk production, about 50% of whey is destined for human consumption. However, depending on locality and economic factors, 20% to 100% of this residue is used for the benefit of chemical and physical characteristics of the soil (ROBBINS; LEHRSCHE, 1998).

The objective of this study was to determine chemical changes and the mobility and distribution of solutes after fractionated application of whey in columns filled with samples of a typical dystrophic Red-Yellow Oxisol (Oxisol) and a and a typical dystrophic fluvic Inceptisol (Inceptisol), withdrawn at two depths. In addition, the purpose of this study was to characterize the percolate from this application.

MATERIAL AND METHODS

In the laboratory, a test was carried out using soil columns in which whey percolation was performed. The treatments corresponded to a 2 x 2 factorial, being samples of a Oxisol and Inceptisol collected at two depths (0-20 and 20-40 cm), in the municipality of Paula Cândido, Zona da Mata of Minas Gerais. The collection sites were selected among those with possibilities of receiving whey applications, considering that hillside areas, represented by Oxisol, and fluvial terraces used for pasture, represented by Inceptisol, are common in the region.

Samples of the 0-20 and 20-40 cm layers of these soil classes were collected air dried and sieved with 2 mm aperture for chemical and physical characterization (Table 1).

Table 1: Chemical and physical characterization of Oxisol and Inceptisol samples.

Determination	Oxisol		Inceptisol	
	0–20 cm	20–40 cm	0–20 cm	20–40 cm
pH in water (1:2,5)	5.23	4.90	6.05	5.75
EC (es) (mS m ⁻¹) ⁽¹⁾	20.8	6.20	29.4	8.80
N total (mg dm ⁻³) ⁽³⁾	1570	980	1250	380
K (mg dm ⁻³) ⁽⁵⁾	54	16	105	113
Na (mg dm ⁻³) ⁽⁵⁾	4.80	1.70	4.80	3.80
Ca ²⁺ (cmol _c dm ⁻³) ⁽⁶⁾	1.30	0.38	2.47	1.18
Mg ²⁺ (cmol _c dm ⁻³) ⁽⁶⁾	0.77	0.22	1.23	0.71
Al ³⁺ (cmol _c dm ⁻³) ⁽⁷⁾	0.30	0.90	0	0.10
P (mg dm ⁻³) ⁽⁸⁾	2.1	1.0	3.4	1.0
Organic matter (g kg ⁻¹) ⁽⁹⁾	49.4	27.9	33.0	12.7
P remaining (mg L ⁻¹) ⁽¹⁰⁾	28.30	20.20	44.70	34.50
Particle size analysis ⁽¹¹⁾				
- Sand (kg kg ⁻¹)	0.433	0.332	0.295	0.177
- Silt (kg kg ⁻¹)	0.076	0.081	0.239	0.328
- Clay (kg kg ⁻¹)	0.491	0.587	0.466	0.495

(1) Electric conductivity in saturation paste extract (Richards, 1954). (2) Electrical conductivity in the 1:5 extract (Richards, 1954). (3) Modified Kjeldahl method (TEDESCO et al., 1985). (4) extractor KCl 1 mol L⁻¹ (TEDESCO et al., 1995). (5) Mehlich¹ extractor (DEFELIPO; RIBEIRO, 1997). (6) KCl extractor 1 mol L⁻¹ (Vettori, 1969). (7) KCl extractor 1 mol L⁻¹ (Vettori, 1969). (8) Extractor Mehlich-1 (BRAGA; DEFELIPO, 1974). (9) Modified Walkley-Black method (DEFELIPO; RIBEIRO, 1997). (10) Alvarez V et al., (2000). (11) Ruiz (2005).

Each experimental unit consisted of soil samples placed in 11 PVC rings, 6.6 cm in diameter and 7 cm high, overlapped and sealed with silicone glue in the joints. At the lower end of the segment, a perforated PVC plug was fitted, with a funnel attached to it, employing plastic mass for sealing. On the last centimeter of the lower ring was placed a thin layer of glass wool and, over it, 1 cm of very thick sand (2.0-1.0 mm), to facilitate drainage. Then another layer of glass wool was placed to avoid mixing the soil with the sand.

The inner walls of the rings received a layer of liquid resin on which was spread a mixture of washed coarse and fine sand in order to avoid preferential flow of water through the walls of the column. Externally, the rings were attached with adhesive tape, to mechanically stabilize the column, which was filled with soil up to 7 cm from the top edge, forming a column of 70 cm height and volume of 2.4 dm³.

The soil, sieved by a 4 mm mesh, was placed with the help of a funnel, making circular movements to prevent the segregation of larger aggregates in the center of the column and, thus, changing the section of the infiltration. In addition, the column was slightly compacted, dropping it vertically from a height of approximately 0.5 cm from the ground three times, thus ensuring the desired soil volume (2.4 dm³).

In order to homogenize the humidity, the soil present in each experimental unit was saturated with deionized water 5 days before the beginning of the test, with the percolated volume discarded. After saturation, the upper end of the column was covered with plastic film to prevent water evaporation.

The treatments were arranged in a randomized complete block design with five replicates. The whey used in the experiment came from the manufacture of fresh minas cheese (Table 2).

Table 2. Physical, chemical and biochemical characterization of whey.

pH ¹	EC ² dS m ⁻¹	BOD ³	COD ⁴	OG ⁵ g L ⁻¹	ST ¹	Total N ⁶	K ⁷	Na ⁷ mg L ⁻¹	Ca ⁸	Mg ⁸
4,73	6,12	37,93	68,11	3,01	67,59	1290	1656	456	555	75

(1) APHA (1995). (2) Electric conductivity (EC) - APHA (1995). (3) Biochemical oxygen demand (BOD)- Winkler method (iodometric) (APHA, 1995). (4) Chemical oxygen demand (COD) - Open reflux method (APHA, 1995). (5) Oils and greases (OG) - Extraction method in Soxhlet (APHA, 1995). (6) Total nitrogen - Modified Kjeldahl method (TEDESCO et al., 1985). (7) Potassium (K) and sodium (Na) - Flame emission photometry. (8) Calcium (Ca) and magnesium (Mg)- Atomic absorption spectrophotometry.

The percolation columns containing the soil received four applications of 0.2 volume of pores of whey each at 6-day intervals. After 24 hours of each application, the percolate was collected and the volume determined. The percolate produced between 24 and 120 hours was also collected. Samples were duly stored under refrigeration for further analysis. The determination of percolates was performed by the modified Kjeldahl method (TEDESCO et al., 1997). With the percolated volume and the concentrations of Na, K, Ca and Mg, total N, N-NH₄⁺, N-NO₃⁻, in the percolated solution, the lost amount of each solute was calculated by leaching.

After 6 days of the last application of whey, the columns were disassembled and the soil contained in the central 5 cm of each ring was removed for analysis. The results of the analyzes were as follows: pH in water, EC (1:5) (RICHARDS, 1954), total N, N-NH₄⁺, N-NO₃⁻ by the modified Kjeldahl method (TEDESCO et al., 1985), Na and K (DEFELIPO; RIBEIRO, 1997) and Ca and Mg (VETTORI, 1969).

The experimental results were analyzed statistically in order to verify the mobility and the distribution of the solutes in the columns of soil and the quantities in the percolated. The dependent

variables were analyzed by means of orthogonal contrasts, to compare soil classes and sampling depths. The results were presented as mean contrasts (ALVAREZ V.; ALVAREZ, 2006). The values determined in the soil column rings were analyzed by regression, using the mean depth as the dependent variable.

RESULTS AND DISCUSSION

The conduction of the experiment in the confined environment of soil columns was complicated due to the appearance of colonies of fungi and bacteria that, together with the accumulation of fat, caused changes in the rate of infiltration in the different leaches. In an attempt to facilitate infiltration, scarification was performed on the upper ring, but the result was not satisfactory. As for leachate collection, there was a marked drop in volume during leaching. This, in fact, could have caused a mixture between the effluents of the last applications. The problems noted led to discarding the soil of the upper ring, submitted to scarification, and to consider only the two initial leaches in the presented results.

To determine the mobility and distribution of solutes in the soil columns, pH, EC (1: 5), total

N, N-NH₄⁺, N-NO₃⁻, K, Na, Ca and Mg were related to the location of each ring. In order to determine regression equations, the dependent variables were analyzed by establishing the mean depth in the ring as an independent variable. Considering the 7 cm length of each ring and that determinations were performed on nine rings, the values of the independent variable were in the range of 3.5 to 59.5 cm.

The application of whey in the studied soils caused a generalized decrease in pH (Table 1 and 3), a response consistent with the acidity of the applied residue (Table 2). The decrease was proportional to the values presented in the characterization (Table 1), registering lower pH in the Oxisol and the samples of the lower layers of the two soils under study.

Table 3: pH, EC (1: 5), total N, N-NH₄⁺, N-NO₃⁻, K, Na, Ca and Mg according to the soil and the sampled layer after four whey leaches.

Attribute	Oxisol		Inceptisol	
	0-20 cm	20-40 cm	0-20 cm	20-40 cm
pH	4.75	4.36	5.19	4.44
EC (1:5) (mS m ⁻¹)	61.8	60.8	71.8	72.0
Total N (mg dm ⁻³)	1997	1336	1681	689
N-NH ₄ ⁺ (mg dm ⁻³)	140	117	123	66
N-NO ₃ ⁻ (mg dm ⁻³)	64	74	61	57
K (mg dm ⁻³)	568	567	611	601
Na (mg dm ⁻³)	143	162	144	146
Ca (cmol _c dm ⁻³)	1.87	1.03	2.83	1.38
Mg (cmol _c dm ⁻³)	0.81	0.32	1.07	0.55

The movement of the whey along the column led to a decrease in pH throughout the depth of the experimental unit (Figure 1). As previously indicated, to the acidity of the product, pH 4.73 (Table 2), oxidation processes of organic matter were added which contributed to this decrease in pH.

The incorporation of whey caused a generalized increase in EC of extract 1:5 (Table 3). Comparing the ECs of the saturation paste extracts and those of the extract 1:5 (Table 1), it is observed

that the ECs determined in the experimental conditions approximate the limit of 4 dS m⁻¹, as indicated by Richards (1954) for differentiate between normal and saline soils. In the comparison between soils, the greatest tendency to salinization occurred in soil Inceptisol, with no difference between layers of the same soil (Table 4). This response can be attributed to the EC of the applied residue, much larger than that of the soils sampled (Tables 1 and 2). With similar mineralogy, the two layers of each soil presented the same behavior.

Table 4: Mean orthogonal contrasts (C) of pH, EC (1:5), total N, N-NH₄⁺, N-NO₃⁻, K, Na, Ca and Mg according to the soil and the sampled layer after four leaches with whey ⁽¹⁾

pH	EC mS m ⁻¹	Total N	N-NH ₄ ⁺	N-NO ₃ ⁻ mg dm ⁻³	K	Na	Ca	Mg cmol _c dm ⁻³
C1	-0.26*	-10.6*	482*	35*	10	-38	7	-0.66*
C2	0.40*	1.0	661*	23*	-10	1	-19	0.85*
C3	0.76*	-0.2	992*	57*	4	10	-2	1.45*

1) C1: Oxisol vs Inceptisol. C2: layer 0-20 cm vs layer 20-40 cm d / Oxisol. C3: layer 0-20 cm vs layer 20-40 cm d / Inceptisol. * Significant at 5% by the F test.

The EC distribution of extract 1:5 is characterized by appreciable leaching of electrolytes along the column (Figure 01). This is confirmed by the values determined at the lower extremities of the columns, always higher than 34.0 mS m⁻¹ (Table 5). The concentration associated with this EC is approximately 3.5 mmol L⁻¹ (RICHARDS, 1954).

Table 5. Regression equations relating pH, EC (1: 5), total N, N-NH₄⁺, N-NO₃⁻, K, Na, Ca and Mg, to soil column depth (P) considering the soil and the layer sampled (LS) after four leaches with whey

Dependent Variable	Soil	LS	Regression Equation	R ²
pH	Oxisol	cm		
		0-20	$\hat{Y} = 4.986 - 0.02706 P + 0.0004682 P^2$	0.937
EC (μS cm ⁻¹)	Inceptisol	20-40	$\hat{Y} = 4.202 + 2.344/P$	0.951
		0-20	$\hat{Y} = 6.904 - 0.7653 P^{0.5} + 0.07465 P$	0.887
	Oxisol	20-40	$\hat{Y} = 10.16 - 3.99 P^{0.5} + 0.797 P - 0.0484 P^{1.5}$	0.962
		0-20	$\hat{Y} = 1606 - 261.3 P^{0.5} + 12.66 P$	0.989
Total N (mg dm ⁻³)	Inceptisol	20-40	$\hat{Y} = 1433 - 45.62 P + 0.4640 P^2$	0.994
		0-20	$\hat{Y} = 1/(0.001002 + 0.00001405 P)$	0.955
	Oxisol	20-40	$\hat{Y} = 630.5 + 449.4 P^{0.5} - 122.8 P + 7.881 P^{1.5}$	0.992
		0-20	$\hat{Y} = 4159 - 745.9 P^{0.5} + 57.03 P$	0.984
N-NH ₄ ⁺ (mg dm ⁻³)	Inceptisol	20-40	$\hat{Y} = 995.5 + 5158/P$	0.952
		0-20	$\hat{Y} = 2940 - 389.1 P^{0.5} + 25.57 P$	0.990
	Oxisol	20-40	$\hat{Y} = 1962 - 378.8 P^{0.5} + 23.41 P$	0.965
		0-20	$\hat{Y} = 516.0 - 129.2 P^{0.5} + 9.843 P$	0.985
N-NO ₃ ⁻ (mg dm ⁻³)	Inceptisol	20-40	$\hat{Y} = 295.9 - 10.53 P + 0.1159 P^2$	0.995
		0-20	$\hat{Y} = 264.6 - 40.51 P^{0.5} + 2.315 P$	0.951
	Oxisol	20-40	$\hat{Y} = 22.58 + 94.82 P^{0.5} - 26.41 P + 1.861 P^{1.5}$	0.970
		0-20	$\hat{Y} = 1/(0.008865 + 0.0002642 P)$	0.809
K (mg dm ⁻³)	Inceptisol	20-40	$\hat{Y} = \bar{Y} = 74$	
		0-20	$\hat{Y} = \bar{Y} = 61$	
	Oxisol	20-40	$\hat{Y} = \bar{Y} = 57$	
		0-20	$\hat{Y} = 2573 - 599.9 P^{0.5} + 37.42 P$	0.994
Na (mg dm ⁻³)	Inceptisol	20-40	$\hat{Y} = 1832 - 70.23 P + 0.7182 P^2$	0.989
		0-20	$\hat{Y} = 1331 - 34.12 P + 0.2686 P^2$	0.994
	Oxisol	20-40	$\hat{Y} = 1997 - 365.3 P^{0.5} + 17.24 P$	0.988
		0-20	$\hat{Y} = 387.4 - 55.89 P^{0.5} + 1.653 P$	0.976
Ca (cmol _c dm ⁻³)	Inceptisol	20-40	$\hat{Y} = 572.2 - 127.1 P^{0.5} + 8.381 P$	0.974
		0-20	$\hat{Y} = 1/(0.003677 + 0.0001268 P)$	0.966
	Oxisol	20-40	$\hat{Y} = 211.8 - 2.094 P$	0.970
		0-20	$\hat{Y} = \bar{Y} = 1.87$	
Mg (cmol _c dm ⁻³)	Inceptisol	20-40	$\hat{Y} = 3.674 - 0.9037 P^{0.5} + 0.06822 P$	0.996
		0-20	$\hat{Y} = 2.256 + 0.01833 P$	0.910
	Oxisol	20-40	$\hat{Y} = 2.001 - 0.3565 P^{0.5} + 0.04052 P$	0.825
		0-20	$\hat{Y} = 0.5220 + 0.009024 P$	0.981
Inceptisol	20-40	$\hat{Y} = 0.8355 - 0.1942 P^{0.5} + 0.01623 P$	0.980	
	0-20	$\hat{Y} = 0.5987 + 0.01508 P$	0.970	
		20-40	$\hat{Y} = 0.5087 - 0.09540 P^{0.5} + 0.01741 P$	0.942

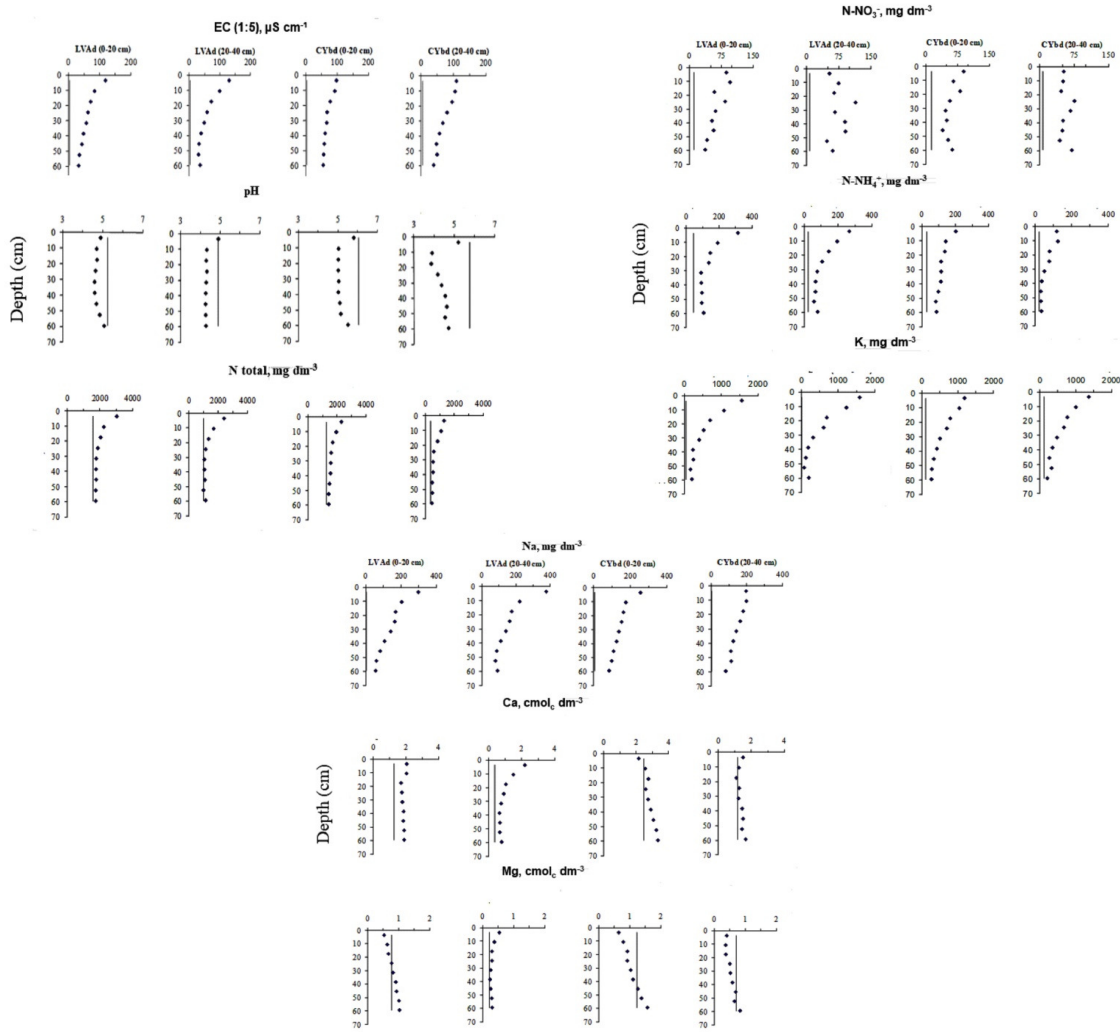


Figure 1. In-depth behavior of: pH, EC (1:5), Total N, N-NO₃⁻, N-NH₄⁺, K, Na, Ca and Mg considering the soil class and the sampling depth. The vertical dash indicates the value in the preliminary characterization of the soil.

Considering the nitrogen compounds (total N, NH₄⁺ and NO₃⁻), it was verified that, in all cases, the application of whey caused the increase of their availability (Tables 1 and 3). However, the total N ratio: N- NH₄⁺: N-NO₃⁻ presented particularities

(Table 6). There was a higher increase in assimilable N (N- NH₄⁺ and N-NO₃⁻) due to the mineralization processes of the organic matter and the higher concentration of these forms in relation to organic N from the incorporated whey (Table 2).

Table 6. Total N ratio: N-NH₄⁺: N-NO₃⁻ in the soil (in the characterization and after four leaching) and applied whey

Material	Oxisol		Inceptisol	
	0-20 cm	20-40 cm	0-20 cm	20-40 cm
Soil				
In the characterization	142.5 : 3.9 : 1	138.4 : 3.0 : 1	87.9 : 2.0 : 1	53.5 : 3.0 : 1
After leaching	31.2 : 2.2 : 1	18.1 : 1.6 : 1	27.6 : 2.0 : 1	12.1 : 1.2 : 1
Whey				
	17,9 : 3 : 1			

There were statistically significant differences for total N and N-NH₄⁺ between soils and between layers within each soil. The greatest increase was in Oxisol and in the upper layers within each soil. The lower levels of N-NO₃⁻ and the lower interaction with the soil ion exchange complex did not show statistically significant differences for this element in the analyzed contrasts (Table 4).

Soil chemical analysis revealed decreases in total N, N-NH₄⁺ and N-NO₃⁻ after application of whey (Table 3). Similar results were found by Gheri et al. (2003), when applying whey on samples of a Red-Yellow Argisol in a degraded pasture area. According to these authors, despite the high doses of N provided to the soil with the application of this residue, there was no significant increase in total N. This fact indicates that the mineralization rate of N compounds was sufficiently fast that they did not allow accumulation of these compounds in the soil.

Considering the mobility and distribution of the nitrogen compounds along the column, the total N applied, composed of a maximum of 18% of N-NH₄⁺ + N-NO₃⁻ (Table 3), the remainder being bound to organic molecules of higher molecular weight and, possibly, lower solubility, recorded appreciable transport up to approximately half of the column (Figure 01). The NH₄⁺, interacting with the cation exchange complex of the soil, moved along the column, but with higher availability in the upper portion (Figure 01). NO₃⁻ was the most mobile nitrogen compound, due to its higher solubility and

lower interaction with the soil particles (Figure 01). Of all the ions studied, it was the only one that did not have adequate adjustment in the regression analysis (Figure 01 and Table 5).

Considering the alkaline and alkaline-earth cations, there is also a general increase in the levels of K, Na, Ca and Mg in the soil after application of whey, in relation to the values determined in the characterization (Tables 1, 3 and Figure 01). However, only statistically significant differences were recorded for Ca and Mg in both the comparison between soils and in the comparison between layers (Table 4). It is observed that the whey had high concentrations of K and Na (Table 2), being found in a lower concentration in the exchange complex, as shown in the soil characterization (Table 1). The characteristic of the displacing fluid led to the Na and K levels close to those of the soil characterization after the four applications, considering that their textural and mineralogical characteristics are similar (Table 1 and 4).

In order to allow comparison, the available K and Na contents and exchangeable Ca and Mg in the previous and subsequent soil leaching characterization (Tables 1 and 3), as well as the concentration of these same elements in the whey (Table 2), were recalculated and their values expressed in cmol_c dm⁻³ and cmol_c L⁻¹, respectively (Table 7). The K: Na: Ca: Mg ratios thus calculated allow us to advance in the discussion of the topic.

Table 7. Levels of K, Na, Ca and Mg in the soil (in the characterization and after four leaches) and applied whey

Material	K	Na	Ca	Mg	K:Na:Ca:Mg
Solo in the characterization (cmol _c dm ⁻³)					
Oxisol (0–20 cm)	0.14	0.02	1.30	0.77	0.18 : 0.03 : 1.69 : 1
Oxisol (20–40 cm)	0.04	0.01	0.38	0.22	0.18 : 0.05 : 1.73 : 1
Inceptisol (0–20 cm)	0.27	0.02	2.47	1.23	0.22 : 0.02 : 2.01 : 1
Inceptisol (20–40 cm)	0.29	0.02	1.18	0.71	0.41 : 0.03 : 1.66 : 1
Soil after leaching (cmol _c dm ⁻³)					
Oxisol (0–20 cm)	1.46	0.62	1.87	0.81	1.80 : 0.77 : 2.31 : 1
Oxisol (20–40 cm)	1.45	0.70	1.03	0.32	4.53 : 2.19 : 3.22 : 1
Inceptisol (0–20 cm)	1.57	0.63	2.83	1.07	1.47 : 0.59 : 2.64 : 1
Inceptisol (20–40 cm)	1.54	0.63	1.38	0.55	2.80 : 1.15 : 2.51 : 1
Whey (cmol _c L ⁻¹)					
	4.25	1.98	2.78	0.63	6.79 : 3.17 : 4.44 : 1

The Ca:Mg ratio, with extremes of 1.69 and 2.01:1 in the soil before the assay was increased to 2.31 and the range 3.22:1 after leaching. From the nutritional point of view, the application of whey, therefore, would not cause imbalance of these two

nutrients, considering that its Ca:Mg ratio is 4.44:1 (Table 7).

Greater attention should be considered regarding K and especially Na. Taking the Mg as a reference, K values between 0.18 and 0.41:1 rose up to 4.53:1. Na, close to 0.03:1 in all cases, increased

to 2.19:1 in the Oxisol, sampled between 20 and 40 cm (Table 7). The Na:Mg ratios were higher in the deeper layers of both soils, indicating greater susceptibility to salinization in samples with lower organic matter content.

As verified by the high ECs in the soil after the application of the whey, it was evidenced the tendency of salinization of these soils with the increase of the whey volume applied. The Na:Mg ratio of this milk residue, 3.14:1, reinforces the need for careful use. On the other hand, the ratio K: Na: Ca: Mg of whey, 6.75: 3.14: 4.41: 1, indicates the possibility of loss of Mg by leaching, by displacement of this nutrient from the exchange, due to the high concentrations of K, Na and Ca (Table 7).

Comparing the transport of K, Na, Ca and Mg in the columns (Figure 01), it is observed that the high concentrations of K, Na and Ca (Table 3) caused Mg displacement of the exchange complex and marked leaching. This is proved by the equation of the curves of this cation (Figure 01) where concentrations lower than those of the characterization in the upper portion of the columns were determined. This response is less visualized for Ca (Figure 01) because of its higher concentration in

whey (Table 2) and the greater interaction with the cation exchange complex of the soil.

Only the first two percolates collected after 24 hours of the application of the whey were analyzed because of the difficulty in the leaching caused by the accumulation of suspended and fat materials, mainly in the upper part of the columns, as previously indicated. Also, in the fourth application of the whey, some columns received inferior volumes than proposed, as a function of the decrease of the infiltration. The application of 20% of the pore volume of whey may have caused organic overload and surface sealing because it was carried out in a reduced time interval (6 days between applications), leading to a decrease in the hydraulic conductivity and a consequent decrease in leached volume (Table 8). This fact indicates that, although there was an increase in microbial activity, a more adequate whey management would be necessary. In this sense, field trials would be satisfactory to prove that longer intervals between applications or whey dilution, if economically feasible, could increase the volume of collected leachate, thus alleviating problems related to low hydraulic conductivity.

Table 8. Volume of the leachate after 24 and 120 hours of application of 0.2 volume of whey pores, according to the soil and the sampled layer. Values in parentheses indicate proportion of the volume leached relative to the volume applied in each leach.

Volume	Oxisol		Inceptisol	
	0–20 cm	20–40 cm	0–20 cm	20–40 cm
	mL			
Applied	296	312	305	315
	First leaching			
Leached 0–24 h	235 (0.794)	243 (0.778)	241 (0.790)	231 (0.733)
Leached 24–120 h	20 (0.068)	35 (0.112)	28 (0.092)	55 (0.175)
Leached total	255 (0.862)	278 (0.890)	269 (0.882)	286 (0.908)
	Second leaching⁽¹⁾			
Leached 0–24 h	215 (0.727)	178 (0.571)	186 (0.610)	232 (0.737)
Leached 24–120 h	23 (0.078)	19 (0.061)	20 (0.066)	31 (0.098)
Leached total	238 (0.805)	197 (0.631)	206 (0.675)	263 (0.835)
	Third leaching⁽¹⁾			
Leached 0–24 h	78 (0.264)	105 (0.337)	180 (0.590)	188 (0.597)
Leached 24–120 h	29 (0.098)	28 (0.090)	25 (0.082)	27 (0.086)
Leached total	107 (0.362)	133 (0.426)	205 (0.672)	215 (0.683)
	Fourth leaching^{(1),(2)}			
Leached 0–24 h	46 (<0.156)	100 (<0.321)	142 (<0.466)	133 (<0.422)
Leached 24–120 h	37 (<0.125)	44 (<0.141)	32 (<0.105)	29 (<0.092)
Leached total	83 (<0.281)	144 (<0.462)	174 (<0.570)	162 (<0.514)

(1) Interval between leaching: 144 hours. (2) Some columns, in this leaching, received volumes lower than indicated, due to the decrease of infiltration.

The determinations carried out in the percolates included pH, EC, COD, total N, N-NH₄⁺,

N-NO₃⁻, K, Na, Ca and Mg. Values different from seven for pH and zero in other determinations

indicate that in all cases there was a leaching of solutes that contributed by the whey and possibly displaced from the soil exchange complex. In the comparison between soils and layers, larger losses of solutes in the Inceptisol and the 0-20 cm layer are

observed in general, compared to the samples taken at 20-40 cm. In leachates there was, in general, a decrease in pH and an increase in solute loss in the second leaching, compared to the first one (Table 9).

Table 9. Mean orthogonal contrasts (C) of pH, COD and total N, N-NH₄⁺, N-NO₃⁻, K, Na, Ca and Mg in the effluent collected in 24 hours in the first two leaches, according to the soil and the sampled layer ⁽¹⁾

Determination	C1	C2	C3	C4	C5	C6	C7
pH	-0.73	-0.88	-0.345	0.55	-0.51	1.58	0.90
EC (dS m ⁻¹)	-0.35	1.03	1.58	-0.69	-0.050	-1.26	-0.16
COD (mg L ⁻¹)	-514	2442	3058	-2152	-460	1865	1035
Total N (mg/column)	1.64	9.89	11.55	-5.95	-2.93	-3.58	-0.41
N-NH ₄ ⁺ (mg/column)	0.97	8.43	9.16	-3.81	0.51	-0.78	0.25
N-NO ₃ ⁻ (mg/column)	0.036	-0.62	0.20	-0.20	-0.82	0.56	0.21
K (mg/column)	VNH ^{2/}	5.51	4.18	-0.77	0.03	-0.85	-2.76
Na (mg/column)	-0.54	1.81	1.26	1.70	-0.19	-1.01	-0.95
Ca (mg/column)	-12.35	20.20	41.47	-10.79	-0.35	-42.09	-2.52
Mg (mg/column)	VNH ⁽²⁾	10.81	10.36	-5.70	0.21	1.45	-1.39

⁽¹⁾ C1: Oxisol vs Inceptisol. C2: Layer 0-20 cm vs. Layer 20-40 cm d / Oxisol. C3: Layer 0-20 cm vs Layer 20-40 cm d / Inceptisol. C4: Compares leachates 1 and 2 d / layer 0-20 cm d / Oxisol. C5: Compares leachates 1 and 2 d / layer 20-40 cm d / Oxisol. C6: Compares leachates 1 and 2 d / layer 0-20 cm d / Inceptisol. C7: Compares leachates 1 and 2 d / layer 20-40 cm d / Inceptisol. ⁽²⁾ VNH: Inhomogeneous variances significant at 5% by the F test.

Table 10 shows the relationships between total N, N-NH₄⁺ and N-NO₃⁻ determined for each soil, sampling depth and leaching. It is observed that, independently of the treatment, the sum of N-NH₄⁺ and N-NO₃⁻ is almost equal to the concentration of total N, revealing that organic nitrogen was almost totally degraded. Any

differences should be attributed to the method of determination. Thus, other nitrogenous compounds were practically not lost by leaching in the columns. The loss of N in its most soluble forms should be taken into consideration when applying whey, due to the potential of this residue.

Table 10. Total N ratio: N-NH₄⁺ and N-NO₃⁻ in the effluent collected in 24 hours in the first and second leaches, according to the soil and the sampled layer

Leaching	Oxisol		Inceptisol	
	0-20 cm	20-40 cm	0-20 cm	20-40 cm
First	10.3 : 8.2 : 1	2.3 : 2.0 : 1	6.3 : 5.7 : 1	1.3 : 0.99 : 1
Second	13.1 : 9.7 : 1	2.7 : 1.1 : 1	11.7 : 8.2 : 1	1.9 : 0.96 : 1

To compare the amount of leached K, Na, Ca and Mg, were transformed to mmol_c, and concentrations were calculated by dividing the amounts by the leached volume (Table 8). The results, together with those from whey characterization (Table 2), all expressed as mmol_c L⁻¹, are shown in Table 11.

In the characterization of whey, the concentration sequence is: K > Ca > Na > Mg. In the set of leachates, Ca > Mg > K > Na.

There are two changes in this sequence when exchanging Na and K position, but without appreciably altering the general trend, ie Ca > Mg and K > Na. The distribution of K, Na, Ca and Mg in the soil columns (Figure 01) shows a distinct trend in the distribution of these cations, with a higher surface concentration for K and Na. The Ca shows little change in depth in the Oxisol, even considering that the added residue is rich in Ca. In

the Inceptisol there is a clear trend of leaching loss. The same can be indicated for Mg.

From the comparison between the values of Tables 11 and Figure 01, it was concluded that K, in higher concentration in the whey, but with greater interaction with the soil particles, was proportionally less leached than Na, found in very low availability in soils and layers in study (Table

1). The two monovalent cations contribute to the displacement of Ca and Mg - the latter in a higher proportion, due to lower concentration in the applied whey and less interaction with the cation exchange complex of the soil.

Table 11. Concentration of K, Na, Ca and Mg in whey and effluent collected in 24 hours in the first and second leaches, according to the soil and the sampled layer

Material	K	Na	Ca	Mg	K : Na : Ca : Mg
	mmol _c L ⁻¹				
Whey	42.462	19.826	27.750	6.250	6.79 : 3.17 : 4.44 : 1
	First leaching				
Oxisol (0–20 cm)	0.660	0.627	3.702	3.071	0.21 : 0.20 : 1.21 : 1
Oxisol (20–40 cm)	0.099	0.113	0.498	0.278	0.36 : 0.41 : 1.79 : 1
Inceptisol (0–20 cm)	1.009	0.415	5.131	4.485	0.22 : 0.09 : 1.14 : 1
Inceptisol (20–40 cm)	0.482	0.201	0.660	0.429	1.12 : 0.47 : 1.54 : 1
	Second leaching				
Oxisol (0–20 cm)	0.813	0.340	6.556	5.566	0.15 : 0.06 : 1.18 : 1
Oxisol (20–40 cm)	0.131	0.200	0.778	0.281	0.47 : 0.71 : 2.77 : 1
Inceptisol (0–20 cm)	1.424	0.774	17.962	5.161	0.28 : 0.15 : 3.48 : 1
Inceptisol (20–40 cm)	0.786	0.379	1.200	0.927	0.85 : 0.41 ; 1.30 : 1

CONCLUSIONS

The application of whey in the soil as the final destination for this residue is an environmentally viable alternative, provided that a correct management is made in order to avoid problems of surface sealing, salinization and leaching of undesirable elements.

The high electrolyte concentrations of whey caused a general increase of EC in the studied soils, suggesting a tendency to salinization.

There was a higher increase in the assimilable nitrogen content (N-NH₄⁺ and N-NO₃⁻) in the soil due to the mineralization processes of the organic matter and the higher concentration of these forms in relation to the total N in the whey.

High concentrations of K, Na and Ca caused Mg displacement of the exchange complex and marked leaching of this nutrient.

The Na: Mg ratio of 3.14: 1 in the whey reinforces the need for careful disposal of this effluent in the soil.

RESUMO: Entre os setores da indústria alimentícia, o segmento de laticínios é um dos mais importantes do Brasil, sendo o soro de leite o seu maior subproduto. Devido às dificuldades de reaproveitamento e aos impactos ambientais causados, quando descartado como efluente em corpos de água, uma alternativa para a sua destinação final seria a aplicação desse resíduo no solo. Objetivou-se com este trabalho determinar alterações químicas e a mobilidade e distribuição de solutos no solo após aplicação fracionada de soro de leite, assim como a caracterização dos percolados recolhidos após cada aplicação. O ensaio, em laboratório, foi conduzido em colunas de PVC preenchidas com solo. Os tratamentos corresponderam a um fatorial 2 x 2, sendo amostras de um Latossolo Vermelho-Amarelo distrófico típico (LVAd) e de um Cambissolo Flúvico Tb distrófico (CYbd), coletados em duas profundidades: 0–20 e 20–40 cm. Cada unidade experimental foi constituída de 11 anéis de PVC, com 6,6 cm de diâmetro interno e 7 cm de altura. As colunas foram dispostas em um delineamento experimental em blocos casualizados, com cinco repetições. Foram realizadas quatro aplicações de soro de leite, correspondentes a 0,2 volume de poros de solo, a cada intervalo de seis dias. O percolado foi recolhido após 24 e 120 horas de cada aplicação sendo realizadas as seguintes determinações: pH, condutividade elétrica (CE), demanda química de oxigênio (DQO), teores de N total, N-NH₄⁺, N-NO₃⁻, Na, K, Ca e Mg. Após seis dias da última aplicação de soro de leite, as colunas foram desmontadas e o solo de cada anel analisado. As análises compreenderam: pH em água, CE (1:5), N total, N-NH₄⁺, N-NO₃⁻, Na, K, Ca e Mg. As elevadas

concentrações eletrolíticas do soro de leite provocaram aumento generalizado da CE no solo. Houve incremento maior do nitrogênio assimilável (N-NH_4^+ e N-NO_3^-) no solo em decorrência de processos de mineralização. Elevadas concentrações de K, Na e Ca causaram deslocamento de Mg do complexo de troca. Conclui-se que a aplicação de soro de leite no solo é uma alternativa viável do ponto de vista ambiental, desde que seja feito um correto manejo, a fim de evitar problemas de salinização e perdas por lixiviação de elementos indesejáveis.

PALAVRAS-CHAVES: Mineralização de nitrogênio. Lixiviação. Salinização.

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