

Quantitative role of mast cells in odontogenic cystic enlargement

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Abstract

Aim: Mast cells have been hypothesized to play a significant role in pathogenesis of odontogenic cysts. The aim of this study was to evaluate mast cell distribution in cystic lining and the capsule to formulate a mechanism of cystic expansion. **Methods:** Ten formalin-fixed paraffin embedded tissue blocks each of OKC, dentigerous and radicular cysts were selected. Toluidine blue staining (1% in 1% NaCl solution) was done in 5µm thick sections and counting performed in 10 areas using an ocular grid. Areas counted were divided into 4 zones: intraepithelial, subepithelial, intermediate and deep zones (Group I, II, III and IV respectively). **Statistical analysis:** Mean ± S.D. was calculated in each group followed by paired 'T' test. **Results:** Mast cells had greatest concentration in subepithelial zone. 'T' test showed no significant differences between group I and II zones in OKC but a highly significant difference between groups I and II in dentigerous cyst. Radicular cysts showed a significant difference between groups II and III. **Conclusion:** Mast cell degranulation releases numerous hydrolytic enzymes that facilitate breakdown of capsular matrix increasing the hydrostatic pressure due to raised osmolality. Influx of tissue fluids results in their enlargement coupled with resorption at the bone-cyst interface.

Key words: mast cells, degranulation, toluidine blue, odontogenic cysts

Introduction

Odontogenic cysts are possibly the most common benign destructive lesions in the human maxillofacial skeleton. Three most common jaw cysts- Radicular cysts, Odontogenic Keratocysts and Dentigerous cysts (of developmental odontogenic origin) are characterized by an expansile non – infiltrative growth, resulting in a smooth and usually unilocular cavity containing fluid or semi fluid material, lined by an epithelium and supported by a fibrous connective tissue capsule¹.

The expansion of the jaw cyst involves destruction of the extra-cellular matrix due to proteolysis of collagen fibers, osteoid – derived gelatin and protein components of basement membrane¹. Mast cells contain numerous cytoplasmic granules, which are degranulated into the extra-cellular space upon activation. In addition to preformed granule contents, activated mast cells can synthesize de novo vasoactive mediators, for example,

platelet – activating factor, chemotactic mediators, and several proinflammatory cytokines such as IL-1 α , IL-3, IL-6 and TNF – α . Furthermore, mast cells are a rich source of heparin and proteolytic enzymes, such as trypsin, chymase and hyaluronic acid, which participate in connective tissue breakdown in the capsule during normal metabolic turnover, as well as in inflammation¹.

Products released by mast cell activation and subsequent breakdown products of connective tissue elements are released into the cyst lumen increasing the hydrostatic pressure with subsequent enlargement.

The aim of this study was to evaluate mast cell distribution in cystic lining and the capsule to formulate a mechanism of cystic expansion using morphometric analysis.

Material and Methods

Paraffin – embedded formalin fixed tissue blocks of Odontogenic Keratocysts, Dentigerous cysts and Radicular cysts (10 each) were retrieved from the archives of the Department of Oral Pathology, Manipal College of Dental Sciences, Mangalore. Sections of 5µm were cut and stained with freshly prepared Toluidine blue solution (1% toluidine blue in 1% sodium chloride), mounted with DPX, and followed by mast cells counting under 40X magnification.

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Counting of mast cells:

Mast cells were counted in ten areas under 40X magnification using an ocular grid with a total area of 0.30625mm², divided in four zones:

1. Intraepithelial
2. Sub-epithelial
3. Intermediate
4. Deep

For intraepithelial counting, the graticule was oriented along the basement membrane and along the connective capsular tissue at the epithelial-capsular junction for counting in subepithelial zone. Every alternate microscopic field was counted. The graticule was then moved further down into two microscopic fields into the capsule and the procedure repeated for intermediate zone. It was then moved further down two microscopic fields into the capsule to a

third level (deep zone) and the counting was performed in similar manner.

Statistical analysis

Means and standard deviation of the mast cell counting were calculated in each layer zone. For comparisons between zones, the *t* test was applied.

Results

Statistical analysis revealed an increase in mast cell count in all the three cysts at the sub-epithelial zone. Paired ‘T’ test showed no statistically significant difference (Table 1) between intraepithelial (I) and sub-epithelial zones (II) in Odontogenic Keratocysts (*p*=0.08), whereas a highly significant difference (Table 2) was noted between the intraepithelial (group I) and sub-epithelial zones (II) of

Table 1 - Mast cell distribution in odontogenic keratocysts (Mean ± standard deviation and *p* values of *t* test between zones)

Tissue blocks	Intraepithelial(I)	Sub-epithelial(II)	Intermediate(III)	Deep(IV)
1	11	96	50	11
2	0	10	7	0
3	5	11	0	0
4	13	14	10	9
5	10	18	13	4
6	0	1	0	0
7	0	22	17	6
8	0	10	7	0
9	2	12	17	21
10	4	11	3	2
Mean±SD	4.5±5.1	20.5±21.6	12.4±14.6	5.3±6.1
<i>t</i> test (<i>p</i> values)	(I)	(II)	(III)	
(II)	0.083	-	-	
(III)	0.123	0.416	-	
(IV)	0.769	0.102	0.180	

Table 2 - Mast cell distribution in dentigerous cysts (Mean ± standard deviation and *p* values of *t* test between zones)

Tissue blocks	Intraepithelial(I)	Sub-epithelial(II)	Intermediate(III)	Deep(IV)
1	1	6	5	0
2	0	3	0	0
3	4	29	28	36
4	3	15	1	0
5	1	1	0	0
6	5	7	0	0
7	2	28	26	0
8	2	9	3	0
9	8	23	10	0
10	0	41	30	24
Mean±SD	2.6±2.5	16.2±13.4	10.3±12.6	6±12.1
<i>t</i> test (<i>p</i> values)	(I)	(II)	(III)	
(II)	0.005			
(III)	0.074	0.324		
(IV)	0.426	0.101	0.462	

Table 3 - Mast cell distribution in radicular cysts (Mean \pm standard deviation and *p* values of *t* test between zones)

Tissue blocks	Intraepithelial(I)	Sub-epithelial(II)	Intermediate(III)	Deep(IV)
1	0	11	2	34
2	0	6	0	0
3	9	4	3	3
4	2	26	6	1
5	1	9	1	0
6	2	11	3	0
7	0	6	2	4
8	1	2	0	0
9	12	16	7	0
10	1	5	1	0
Mean \pm SD	2.8 \pm 4.2	9.6 \pm 7.1	2.5 \pm 2.4	4.2 \pm 10.6
<i>t</i> test (<i>p</i> values)	(I)	(II)	(III)	
	(II)	-	-	
	(III)	0.008	-	
	(IV)	0.196	0.626	

Dentigerous cysts ($p=0.005$). On the other hand, Radicular cysts showed a highly significant difference (Table 3) between sub-epithelial (II) and intermediate (III) zones ($p=0.007$).

A significant elevation in mean \pm SD values was noted in mast cell population in Odontogenic Keratocysts in the subepithelial zone as compared to Dentigerous and Radicular cysts.

1. Intraepithelial Sub-epithelial Intermediate Deep

Discussion

Mast cells are found widespread throughout the connective tissue wall of all the cysts particularly in the subepithelial zone and are source of a variety of proteolytic enzymes found in the cystic fluid.

The results of the present study showed a great concentration of mast cell in subepithelial zone in all cyst walls. This concentration was higher in OKCs than dentigerous and radicular cysts, suggesting an increased breakdown of capsular matrix in OKCs. OKC epithelium has been shown to be nonkeratinized at places, which causes a transport of breakdown matrix products into the cystic lumen², and consequently can determine an elevated osmolality of the cystic fluid, which partly explains the greater aggressiveness of OKC comparing to other odontogenic cysts. Smith et al.¹ found a considerable amount of mast cells in the walls of odontogenic keratocysts, dentigerous and radicular cysts with the highest concentration seen in sub-epithelial zone. Mast cells were also observed in the epithelial linings, which the authors suggested to be due to a chemotactic stimulus attracting them to the epithelial lining or luminal fluid contents. Smith et al.² concluded that the major source of glycosaminoglycans and proteoglycans in cystic fluid was from the ground substance of the connective tissue capsule,

released because of normal metabolic turnover and inflammatory degradation. Degranulating mast cells release heparin and other hydrolytic enzymes, which facilitate breakdown of glycosaminoglycans and proteoglycans^{2,3}. Histochemical investigations of the connective tissue capsule in odontogenic cysts have demonstrated that hyaluronic acid, a product of mast cell degranulation, is the predominant glycosaminoglycan present along with less amounts of sulphated glycosaminoglycans^{2,3}. The release of glycosaminoglycans and proteoglycans into the luminal fluid contributes significantly to osmotic and hydrostatic pressure by increasing the osmolality of the cyst fluid, thereby raising the internal hydrostatic pressure^{2,3}. Cyst expansion is also affected by the rate in which the surrounding bone is destroyed particularly at the cyst-bone interface⁴.

Teronen et al.⁴ stated that activated mast cells can synthesize vasoactive and chemotactic mediators (e.g., platelet – activating factor) as well as several pro-inflammatory cytokines such as IL-3, IL-6 and TNF- α de novo. These chemical mediators increase vascular permeability thereby facilitating influx of highly osmolar substances in cystic lumen. The highest concentration of mast cells in OKCs explains a greater expansion as compared to other odontogenic cysts. The authors also found a high number of extensively degranulated mast cells in the area of cyst expansion at the border with the bony wall suggestive of high activity of mast cells in this area.

Mast cell degranulation also releases tryptase and prostaglandins which aid in bone resorption which is a feature in cyst enlargement at cyst-bone interface. In addition, interleukin-1 α in OKC cyst wall has been found to have an enhancing effect on matrix metalloproteinases secreted by fibroblasts⁵.

Several other studies have also substantiated the effect of MMPs along with tissue inhibitor of metalloproteinases and collagenases in cyst walls^{6,7}. These cell products have been found to be stimulated by mast cell derivatives⁷⁻⁹. Based upon the literature review analysis, it can be proposed that the degranulating mast cells release products that contribute to cystic enlargement in four ways:

1. By direct release of heparin in luminal fluid
2. By release of hydrolytic enzymes which degrade capsular extra- cellular matrix components thereby facilitating their passage into the fluid
3. By the action of histamine on smooth muscle contraction and vascular permeability encouraging translation of serum proteins
4. By stimulating the production of prostaglandins, interleukin-1 α , TIMP and other collagenases, which are said to be important in bone resorption and thus, cyst growth¹⁰

Based upon the present study and similar investigations, it can be concluded that mast cells play a vital role in the pathogenesis of odontogenic cysts as an elevated number of mast cells was found in the connective tissue capsule of all three odontogenic cysts. The luminal fluid which accumulates as a result of osmolar concentration of mast cell by-products plays an important in cyst enlargement.

References

1. Smith G, Smith AJ, Basu MK. Mast cells in human odontogenic cysts. *J Oral Pathol Med.* 1989; 18: 274 – 8.
2. Smith G, Smith AJ, Browne RM. Histochemical studies on glycosaminoglycans of odontogenic cysts. *J Oral Pathol.* 1988; 17: 55-9.
3. Smith G, Smith AJ, Browne RM. Glycosaminoglycans in fluid aspirates from odontogenic cysts. *J Oral Pathol.* 1984; 13: 614-21.
4. Teronen O, Hietanen J, Lindqvist C, Salo T, Sorsa T, Eklund KK, et al. Mast cell-derived tryptase in odontogenic cysts. *J Oral Pathol Med.* 1996; 25: 376 – 8.
5. Kubota Y, Oka S, Nakagawa S, Shirasuna K. Interleukin-1 α enhances type I collagen-induced activation of matrix metalloproteinase-2 in odontogenic keratocyst fibroblasts. *J Dent Res.* 2002; 81: 23-7.
6. Docherty AJ, Murphy G. The tissue metalloproteinase family and the inhibitor TIMP: a study using cDNAs and recombinant proteins. *Ann Rheum Disease.* 1990; 49: 469-79.
7. Teronen O, Salo T, Laitinen J, Tornwall J, Ylipaavalniemi P, Kontinen YT et al. Characterization of interstitial collagenases in jaw cyst wall. *Eur J Oral Sci.* 1995; 103: 141-7.
8. Taylor AC. Collagenolysis in culture tissue II. Role of mast cells. *J Dent Res* 1971; 50: 1301-6.
9. Suzuki K, Lees M, Newlands GFJ, Nagase H, Woolly DE. Activation of precursors for matrix metalloproteinases 1 (interstitial collagenases) and 3 (stromelysin) by rat mast cell proteinases I and II. *Biochem J* 1995; 305: 301-6.
10. Mundy GR. Cytokines and local factors which affect osteoclast function. *Int J Cell Cloning* 1992; 10: 215-22.