

ROMANIAN
NEUROSURGERY

Vol. XXXIV | No. 3 September 2020

Cerebrospinal fluid dynamics with its
surgical implications

Harold E. Vasquez,
Yeider A. Durango-Espinosa,
Ezequiel Garcia-Ballestas,
B.V. Murlimanju,
Andrei Fernandes Joaquim,
Luis Rafael Moscote-Salazar,
Amit Agrawal



Cerebrospinal fluid dynamics with its surgical implications

Harold E. Vasquez¹, Yeider A. Durango-Espinosa²,
Ezequiel Garcia-Ballestas², B.V. Murlimanju²,
Andrei Fernandes Joaquim⁴,
Luis Rafael Moscote-Salazar², Amit Agrawal⁵

¹ Faculty of Medicine, University of Sinu, Cartagena, COLOMBIA

² Center for Biomedical Research (CIB). Faculty of Medicine - University of Cartagena, Cartagena, COLOMBIA

³ Department of Anatomy, Kasturba Medical College, Manipal Academy of Higher Education, Manipal, Karnataka, INDIA

⁴ Prof. of Neurosurgery. University of Campinas (Unicamp), Campinas-SP, BRAZIL

⁵ Department of Neurosurgery, All India Institute of Medical Sciences, Saket Nagar, Madhya Pradesh, INDIA

ABSTRACT

Cerebrospinal fluid (CSF) is largely (70-80%) produced by the choroids plexus of the ventricles and is considered as the plasma ultrafiltrate. While CSF formation, circulation, and composition appear to be physiological and physical, its absorption appears to be mainly physical. The formation, composition, circulation, absorption, and changes in pathological conditions of CSF are discussed briefly in this review article. The CSF pressure dynamics studies provide information about the tightness, elastance, or outflow resistance of the CSF in the CNS. We believe that the present study shall help to provide essential details of CSF physiology which are important to many disciplines including radiology, neurology, and neurosurgery.

INTRODUCTION

Thorough knowledge of CSF dynamics is essential for understanding the intracranial -intraspinal changes due to pathologic conditions. Perhaps, the first time described was by Dandy in 1919 (1). However, years later three components of CSF dynamics was established by Czosnyka et al, wich are CSF formation, circulation, and composition. This, appears to be physiological and physical, and its absorption may be mainly physical. Since most of the surgical treatments involve manipulating the physical principles, it is important to understand the studies, which are emerged on the dynamics of CSF (2). Studies in patients with hydrocephalus and traumatic brain injury has contributed to understand the strong association between CSF dynamics, physical

Keywords
cerebrospinal fluid,
hydrocephalus,
ventricles



Corresponding author:
Luis Rafael Moscote-Salazar

Center for Biomedical Research
(CIB). Faculty of Medicine - University
of Cartagena. Cartagena. Colombia

rafaelmoscote21@gmail.com

Copyright and usage. This is an Open Access article, distributed under the terms of the Creative Commons Attribution Non-Commercial No Derivatives License (<https://creativecommons.org/licenses/by-nc-nd/4.0/>) which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is unaltered and is properly cited.

The written permission of the Romanian Society of Neurosurgery must be obtained for commercial re-use or in order to create a derivative work.

ISSN online 2344-4959
© Romanian Society of
Neurosurgery



First published
September 2020 by
London Academic Publishing
www.lapub.co.uk

and surgical components (3). The main objective of this review is to review CSF dynamics and the association with surgical components.

PHYSIOLOGY OF CSF DYNAMICS

Physiological principles include vascular, neuro-humoral, and sub-cellular regulation, while physical principles include all the fluid dynamics. Some of the well-known physiological principles include Cushing's reflex, which affects the cerebrovascular system. Pressure on ventricular walls affects the secretion of CSF at the aquaporin level of the choroid plexus. Sco-spondin, the peptide secreted by the SCO (subcommissural organ) will regulate the flow of CSF across the aqueduct of Sylvius. However, Sco-spondin is rudimentary in humans. There are approximately 150 ml of CSF, with a rate of 0.3-0.6 ml/min of production in an adult, distributed between 120 and 30 ml in cranial and spinal subarachnoid spaces, with daily volume produced between 500-600 ml (4). The extracellular space is about 15% of the brain volume and about 1500 ml of adult intracranial space consists of 1100 ml of intracellular space, 200 ml extracellular space, 140 ml of cerebrospinal fluid, and about 60 ml of blood at a given time (5). The choroid plexus in the ventricles is the main production site of CSF formation. However, there is a strong association between resistance of CSF and formation, which in many studies has shown to be the first parameter to consider in patients with intracranial hypertension (6-8). Perhaps, Large amounts of CSF are drained primarily into the blood through the arachnoid villi by penetrating the sinus, mainly into the superior sagittal sinus. Understanding this process, may contribute to be the key to comprehend surgical components. As surgery component, according to Monro-Kellie theory; a disbalance between CSF reabsorption and resistance may be the key to develop every illness related to CSF dynamics (9,10).

PHYSIOLOGICAL RESPONSES

Hypothermia decreases the CSF volume by 11%, with each one-degree reduction in the temperature and hyperthermia increases the CSF volume. In general, the drugs that increase the cerebral blood flow consequently increase the CSF production; on the other hand, the aging process and some infections cause reduce in the CSF formation (11-14). However, the total CSF volume increases with age in both

sexes, mainly due to the contribution by the cortical sulcal volume. These pressure gradients which are being created by continuous CSF secretion and facilitated by the arterial pressure pulsations have an important role for CSF circulation and absorption through the venous system (15). The respiratory variations and vascular pulsations emanating from the choroid plexus and cerebral arteries cause the ventricles to pulsate providing the additional movement to CSF. Although trans-mural migration of CSF across the pial arteries has been demonstrated in the animals, these do not contribute to the bulk of CSF absorption (16).

FUNCTIONS OF CSF

This blood- CSF barrier maintains a chemically precise environment, which is necessary for the neurotransmitter and removes the metabolic products, unwanted drugs and pathological substance that may result in CNS injury (17,18). Changes in CSF Ca, Mg, K produce changes in the sympathetic and autonomic response, ventilation, muscle tone, and emotional state. In continuity with the brain interstitial fluid, CSF provides a stable supply of substrates, primarily glucose, even though the plasma concentration of the substrate is continuously changing. Transport of endorphins, hormones of the hypothalamus and pineal gland is facilitated by the CSF. Specialized ventricular cells and brain parenchymal neurons secrete neuroendocrine factors (19,20).

LABORATORY INVESTIGATION OF CSF

Important information on CSF can be derived from the following parameters, opening pressure, gross appearance, total and differential cell count, bacterial culture and sensitivity, protein and glucose, analysis of immunoglobulins (to detect chronic CNS inflammatory conditions) and cytology (to detect malignant cells). Increased neutrophils in the CSF indicate bacterial meningitis (21). Other causes of increased neutrophil count include a cerebral abscess, seizures, and CNS hemorrhage. Increased lymphocytes in the CSF indicate viral meningitis. Lymphocyte counts are also elevated in meningitis, tuberculosis, syphilis, fungal and parasitic infections. Increased plasma cells is a feature of TB meningitis and chronic inflammatory disorders like multiple sclerosis. The presence of leukemic cells in the CSF indicates meningeal infiltration by leukemic cells (22).

Leukemic cells typically appear in the CSF after several remissions have been achieved by chemotherapy. Tumor cells can be detected by cytological studies and the sources of tumor cells can be from primary CNS tumors like medulloblastoma, or metastatic CNS tumors from lung, breast, GI tract, and melanoma. It may be necessary to determine if a nasal or ear fluid is found as a content in the CSF. CSF fluid contains a modified transferrin protein called β 2-transferrin (tau protein), which is not present in plasma or other fluids. The presence of β 2-transferrin in a fluid strongly suggests that the fluid is CSF, which may be used, by instance, to identify CSF leak after sinus surgery (22,23).

STUDIES OF CSF DYNAMICS

In clinical practice, external lumbar drainage (Tap-test) is widely used to evaluate normal pressure hydrocephalus (NPH). Infusion tests, bolus injections, and isotope dilution methods are employed most often in experimental studies. ICP monitoring in acute conditions such as head injury and chronic conditions such as NPH is widely used. In the study on CSF dynamics by Ramesh et al. a simple, safe, and cost-effective method of infusion method to predict the VPS effectiveness was devised (24). Not only in NPH and PTH but also postmeningitic hydrocephalic patients seems to correlate and benefit from the pre-operative saline infusion studies. CT cisternography consists of the injection of radioactive agents into the subarachnoid space and serial CT scans are taken at different times depending on the clinical problem, such as CSF fistulae, NPH, etc. It is also used to evaluate the shunt function. It can also help in the evaluation of patients receiving intrathecal chemotherapy, evaluation of brain fluid-filled structures (arachnoid cysts, etc.), evaluation of CSF pumps for continuous delivery of medication. CT and MRI are useful to estimate intracranial compliance (25).

CONCLUSION

This review briefs about the formation, composition, circulation and absorption of the CSF. The changes in CSF due to pathological conditions are highlighted. It is believed that the present study will offer the details about the physiology and dynamics of CSF, which is essential to the broad specialties including radiology, neurology, and neurosurgery.

REFERENCES

1. Dandy WE. Internal Hydrocephalus. An Experimental, Clinical and Pathological Study. *Ann Surg* [Internet]. 1919;70(2):129–42. Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1410318/pdf/annsurg00745-0001b.pdf>
2. Czosnyka M, Czosnyka Z, Momjian S, Pickard JD. Cerebrospinal fluid dynamics. *Physiol Meas*. 2004;25(5).
3. Johanson CE, Duncan JA, Klinge PM, Brinker T, Stopa EG, Silverberg GD. Multiplicity of cerebrospinal fluid functions: New challenges in health and disease. *Cerebrospinal Fluid Res*. 2008;5:1–32.
4. Sakka L, Coll G CJ. Anatomy and physiology of cerebrospinal fluid. *Eur Ann Otorhinolaryngol Head Neck Dis*. 2011;128:309–16.
5. Orešković D, Radoš M, Klarica M. Role of choroid plexus in cerebrospinal fluid hydrodynamics. *Neuroscience*. 2017;354:69–87.
6. Bradbury MW, Cserr HF WR. Drainage of cerebral interstitial fluid into deep cervical lymph of the rabbit. *Am J Physiol*. 1981;240:329–36.
7. Pollay M. The pathophysiology of the cerebrospinal fluid circulation. *Neurosurgery, Sci basis Clin Pract* (eds Crockard A, Hayward R, Hoff JT) Blackwells Sci Publ Oxford. 1985;279–96.
8. Linninger AA, Tsakiris C, Zhu DC, Xenos M, Roycewicz P, Danziger Z, et al. Pulsatile cerebrospinal fluid dynamics in the human brain. *IEEE Trans Biomed Eng*. 2005;52(4):557–65.
9. Segal MB PM. The secretion of cerebrospinal fluid. *Exp Eye Res*. 1977;127–48.
10. Klarica M, Orešković D. Enigma of cerebrospinal fluid dynamics. *Croat Med J*. 2014;55(4):287–90.
11. Brierley JB; Field EJ. The connexions of the spinal subarachnoid space with the lymphatic system. *J Anat*. 1948;82:153–66.
12. Welch K; Pollay M. The spinal arachnoid villi of the monkeys *Cercopithecus aethiops sabaeus* and *Macaca irus*. *Anat Rec*. 1963;145:43–8.
13. Cutler RW; Page L; Galicich J; Watters GV. Formation and absorption of cerebrospinal fluid in man. *Brain. a J Neurol*. 1968;91:707–20.
14. Ahmed A; Hickey SM; Ehrett S. et al. Cerebrospinal fluid values in the term neonate. *Pediatr Infect Dis J*. 1996;15:298–303.
15. KC B. Cerebrospinal fluid pressure. *J Neurol Neurosurg Psychiatry*. 1970;33:387–97.
16. Millhorat TH; Hydrocephalus and the cerebrospinal fluid. Williams & Wilkins. 1972;
17. Millhorat TH. The third circulation revisited. *J Neurosurg*. 1975;42:628–45.
18. Sahar A; Hockwald GM; Ransohoff J. Cerebrospinal fluid and cranial sinus pressures. Relationship in normal and hydrocephalic cats. *Arch Neurol*. 1975;23:413–8.
19. Leusen I. Regulation of cerebrospinal fluid composition with reference to breathing. *Physiol Rev*. 1972;52:1–52.
20. Nakada T, Kwee IL. Fluid Dynamics Inside the Brain

Barrier: Current Concept of Interstitial Flow, Glymphatic Flow, and Cerebrospinal Fluid Circulation in the Brain. *Neuroscientist*. 2019;25(2):155–66.

21. Shapey J, Toma A, Saeed SR. Physiology of cerebrospinal fluid circulation. *Curr Opin Otolaryngol Head Neck Surg*. 2019;27(5):326–33.
22. J. SKYPR. ural sinus pressure. In normal and hydrocephalic dogs. *Arch Neurol*. 1964;10:575.580.
23. Greitz D; Hannerz J; Rahn T; Bolander H; Ericsson A. MR imaging of cerebrospinal fluid dynamics in health and disease. On the vascular pathogenesis of communicating hydrocephalus and benign intracranial hypertension. *Acta Radiol*. 1994;35:204–11.
24. Wasserstrom WR; Glass JP; Posner JB. Diagnosis and treatment of leptomeningeal metastases from solid tumors: experience with 90 patients. *Cancer*. 1982;42:759–72.
25. Ramesh VG; Narasimhan V; Balasubramanian C. Cerebrospinal fluid dynamics study in communicating hydrocephalus. *Asian J Neurosurg*. 2017;12:153–8.