

Review article

## Trefoil Factor Family (TFF): Peptides with Numerous Functions

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### Abstract

The trefoil factor family (TFF) consists of a group of small peptides and is highly expressed in tissue that contain mucus-producing cells, predominantly in the mucosa that lines the gastrointestinal tract. Those peptides, which are highly important for epithelial restitution, may act in ways other than using the usual factors responsible for restitution. It was observed that several mechanisms are involved in the TFFs' promotion of restitution. In addition to that, peptides have other functions as well, e.g. they interact with the immune system. Although the TFFs' therapeutic effects have been studied, it is uncertain which of the TFFs' in vitro properties are directly involved when it comes to their in vivo engagement. Observing mice with genetic deletion of TFF peptides can help us discover the function of the peptides that could be indicated by the deletion of the target protein or by adaptive regulation of some other protein that is affected by the deleted gene product. At the very least, a subset of functional networks controlled by a TFF isoform and its downstream effectors can be identified by observing such mice. The discoveries related to the signaling mechanisms of the TFF family leave much to discover about the distinct and shared pathways among those protective peptides.

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## Biological Properties of TFF

The trefoil factor family (TFF) includes a group of small peptides, the first member of which was discovered about thirty years ago (1). TFF1 (formerly known as breast cancer-associated peptide pS2), TFF2 (formerly spasmolytic polypeptide SP) and TFF3 (formerly intestinal trefoil factor) are three members of the trefoil factor family (TFF) known in mammals. These three proteins are small compact peptides that have one or two trefoil domains. While TFF1 and TFF3 contain only one trefoil domain, TFF2 contains two trefoil domains. The basic elements of a trefoil domain are 42–43 amino-acid residues. Six cysteine residues form three disulfide bonds, creating a characteristic three-leafed structure (2).

There are several studies that suggest that TFFs can be regulated by cytokines and transcription factors (especially NF- $\kappa$ B) related to the immune system and that TFFs can regulate them in return, but there is also data suggesting otherwise (3-6).

TFFs have been extensively studied in vivo and in vitro, with most data suggesting that these small peptides improve epithelial repair in the gastrointestinal (GI) tract and other body systems (7).

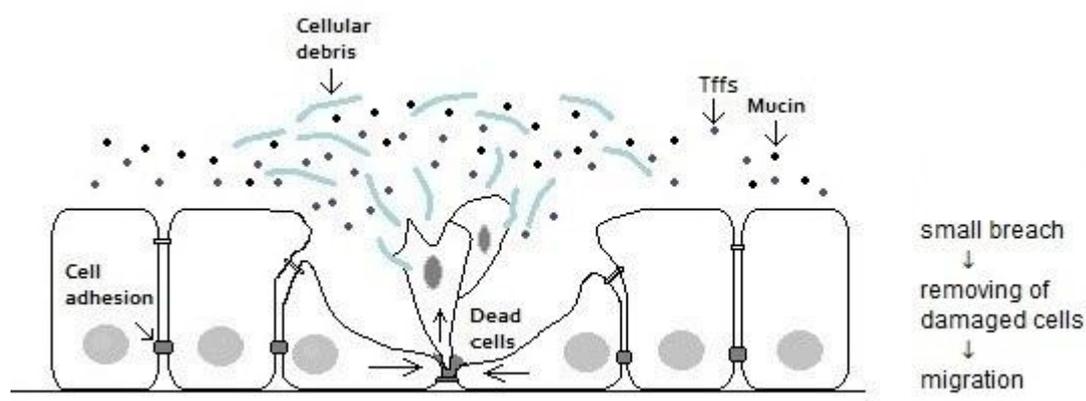
## TFF's Role in Mucosal Protection and Repair

Due to their compact structure, TFFs are relatively resistant to proteolytic degradation in the stomach and small intestine (8). TFFs can mostly be found as secreted molecules in the mucus covering normal epithelium. The predominant site of TFF synthesis is the mucin-producing or goblet cells dispersed in epithelia. All three TFFs are expressed in the stomach, being localized to the surface gastric mucosal cells (9). Their mRNA has been found in the brain,

lungs, trachea, thyroid gland, salivary glands, prostate, uterus and other organs (10). The genes that code them are located on chromosome 21 (11).

Despite the fact that they have been found in almost all tissue containing mucus-secreting cells, TFFs are predominantly expressed in the gastrointestinal tract. Considering their appearance in mucosal tissue, it can be concluded that their functioning might be related to that of mucins. Nevertheless, TFF2 expression is not so common and there is a possibility that different TFFs have different roles in the protection of the epithelium, which is corroborated by the complementary expression of TFFs in the GI tract and by the simultaneous occurrence of each of them with their unique mucin type (MUC); TFF1 appears with MUC5AC, TFF2 with MUC6, and TFF3 with MUC2 (12-14), although gastric and ocular co-localization of TFF3 with MUC5AC also occurs (15, 16). It is speculated that mucosal defense is improved by direct interaction of TFF peptides and mucins.

The role of TFF peptides in cell migration was observed in several studies (17-20), predominantly as a consequence of the response of a damaged epithelium that strives to restore its continuity. In cases of small discontinuity of the epithelium, where cell proliferation is not required, restitution of the epithelium takes place soon after the injury, with coordination of the removal of damaged cells and the migration of healthy epithelial cells into the injured location (Figure 1). The importance of efficient restitution from the physiological viewpoint is high, as the loss of fluids and electrolytes has to be stopped and the luminal antigens and bacteria have to be prevented from entering the tissue and immune cells of the host. Proliferation occurs instead of restitution when tissue is more severely damaged.



**Figure 1. The role of TFF peptides in epithelial restitution**

## TFF and Energy Metabolism

The influence of trefoil factor proteins on energy metabolism can be observed in mice with TFF3 gene knockout (TFF3<sup>-/-</sup> mice). The TFF3 knockout mice have a different expression of miRNA associated with the glycolysis and gluconeogenesis metabolic pathways compared to wild-type mice. The TFF3 knockout mice have a significantly lower body weight compared to the wild type (21). A change in the body mass of mice did not occur in the research with increased expression of the TFF3 gene (22). The fatty changes in the liver of mice have been connected with the change of expression of the TFF3 gene (23). Research has shown that the TFF3 protein participates in glucose metabolism. Similarly, a study showed that hepatic TFF3 expression levels were lower in obese (ob/ob) and high-fat-diet-induced obese mice. Cellular glucose output in mice decreased as a consequence of overexpression of TFF3 in primary mouse hepatocytes, which inhibited the expression of gluconeogenic genes. Experiments using the glucose tolerance test and insulin tolerance test showed that adenovirus-mediated overexpression of TFF3 in diabetic or obese mice improved glucose tolerance and insulin sensitivity. The results also showed that TFF3 peptides are a factor in glucose homeostasis and insulin sensitivity. Consequently, it was concluded that said peptide might be a part of successful modern therapies aimed at metabolic disorders related to type 2 diabetes mellitus (24). Increasing

concentrations of glucose and insulin treatment boosted the expression of TFF3 in intestinal epithelial cells. In addition to that, insulin treatment caused the upregulation of human sodium/glucose cotransporter 1 (hSGLT1), which additionally increased intracellular glucose levels. Downregulation of TFF3 was observed in diabetes mellitus type 1 patients, but the values were modified by insulin treatment. It was discovered that insulin signaling was important for the optimal expression of TFF3 in intestinal epithelial cells, as it elevates intracellular glucose levels and mediates gene expression (25).

Aberrant energy metabolism in the liver promotes insulin resistance, diabetes, and nonalcoholic fatty liver diseases (26). It was recently shown that liver triglyceride accumulation does not cause cellular injury in the liver; the primary causes of liver injury via increased oxidative stress are free fatty acids or their metabolites (27). Changes in lipid metabolism, especially the increase of saturated fatty acids, are associated with increased endoplasmic reticulum (ER) stress, oxidative stress and liver injury in the course of development of fatty liver disease (28). Sirtuin 1 (SIRT1) plays a key role in metabolic regulation, adaptation and oxidative stress. Acting as a nuclear metabolic sensor and deacetylating a wide range of targets, it leads to epigenetic modifications of histones and modulation of transcription factors or metabolic enzymes (29). In addition to SIRT1, peroxisome proliferator-

activated receptors (PPARs) also have an important role in cell metabolism (30).

In case of TFF3 deficiency, the profile and accumulation of fatty acids (FAs) in the liver are affected (Table 1), with no obvious oxidative stress increase, although the expression/activity of monitored enzymes changes, as does the

level of SIRT1 and PPAR $\gamma$  protein. Due to the strong downregulation of hepatic TFF3 in diabetic/obese mice, its presence in circulation and its regulation by food/insulin, TFF3 represents an interesting new candidate for research in metabolic relevant conditions (31).

**Table 1. Fatty acids in liver of Tff3 -/- mice compared to wild type (elevated  $\uparrow$ , decreased  $\downarrow$ )**

| Fatty acids                                    | Tff3 -/-     |
|--|--------------|
| SATURATED                                      |              |
| C14:0 myristic acid                            | $\downarrow$ |
| C18:0 stearic acid                             | $\uparrow$   |
| C20:0 arachidic acid                           | $\uparrow$   |
| MONOUNSATURATED                                |              |
| C16:1 palmitoleic acid ( $\omega$ -9)          | $\downarrow$ |
| C18:1 oleic acid ( $\omega$ -9)                | $\downarrow$ |
| C18:1 vaccenic acid ( $\omega$ -7)             | $\downarrow$ |
| C20:1 eicosenoic (gondoic acid) ( $\omega$ -9) | $\downarrow$ |
| POLYUNSATURATED                                |              |
| C20:2 eicosadienoic acid ( $\omega$ -6)        | $\uparrow$   |
| C20:4 (AA) arachidonic acid ( $\omega$ -6)     | $\uparrow$   |
| C18:3 (ALA) alpha linolenic ( $\omega$ -3)     | $\downarrow$ |
| C22:6 (DHA) docosahexaenoic ( $\omega$ -3)     | $\uparrow$   |
| RATIO $\omega$ -3/ $\omega$ -6                 | $\uparrow$   |

## TFF's Participation in Defense Against Harmful Agents

Another role of the TFF3 protein is the defense of the organism against harmful agents. Mice that cannot synthesize enough TFF3 protein in their liver are deprived of the protective effect of the TFF3 protein in the serum after myocardial (32) and brain ischemia, which consequentially leads to greater tissue damage. Thus, in such mice, a significantly higher activity of caspase 3 and a higher level of cell death in the ischemic cerebral lesion were observed, together with a larger fraction of cerebral infarcts and a smaller fraction of injuries in the cerebral hemisphere, accompanied by more severe forelimb motor deficits. Since the mice were TFF3-deficient, recombinant TFF3 was administered intravenously and it reversed changes in

cerebral injury and forelimb motor function, pointing at the existence of an endocrine neuroprotective mechanism that uses TFF3 from the liver in experimental cerebral ischemia/reperfusion injury (33). TFF3-/- mice have difficulties with regeneration of the mucous membrane of the gastrointestinal tract (34).

High-salt diet (HS) causes endothelial dysfunction and vitiates vascular reactivity to various stimuli. In a recent study, transgenic TFF3-/- mice were introduced as a new model, characterized by a favorable ratio of  $\omega$ -6/ $\omega$ -3 free fatty acids and modified metabolism of arachidonic acid (AA). The results showed that acute HS intake has a much smaller impact on FIR (flow-induced response) in TFF3-/- mice compared to the wild type (WT) (35). The study

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showed that HS intake does not affect NO production in TFF3<sup>-/-</sup> mice (36).

According to another study, although the TFF3 peptide is not expressed in an intact corneal epithelium, its expression is extensively upregulated following an epithelial injury. In addition to that, corneal injuries in TFF3<sup>-/-</sup> mice take much more time to re-epithelialize compared to similar injuries in wild-type mice. In case of alkali-induced corneal wounds, external application of recombinant TFF3 to the wounds speeds up the *in vivo* and combined *in vivo/in vitro* model wound healing in both wild-type and TFF3<sup>-/-</sup> mice. This proves that TFF3 has a key role in the mechanism of corneal wound healing, which opens a possibility of creating new ways of coping with non-healing wounds (37).

### **TFF in the Respiratory System, Pregnancy and Tumorigenesis**

A study (38) describing a murine asthma model found that trans-differentiating Clara cells specifically express TFF1 which is stored in a specific subset of secretory granules. This is proof that TFF1 is an autocrine factor for the trans-differentiation of Clara cells into goblet cells. Another study (39) showed that TFFs play such a role in the differentiation of the airways as well, showing the induction of TFF3 synthesis with the differentiation in *in vivo* humanized tracheal xenograft and *in vitro* air-liquid interface culture models. In addition to that, exogenous TFF3 promoted differentiation of ciliated cells in an EGF-receptor-dependent manner. Both studies implied that TFFs may have important roles in different processes of differentiation of airways, making them promising new targets in treatment of severe chronic and acute airway diseases.

Dynamic changes of trefoil factor proteins in a pregnant woman's serum point at their importance in embryogenesis (40). The presence of the TFF3 protein in the cartilage of mice fetuses during endochondral ossification has been identified, while the exclusion of the TFF3 gene causes changes to the

histomorphological structure of cancellous bone, as well as hearing disorders and accelerated presbycusis, which indicates that it has a role in morphogenesis of organs (41-43).

The TFF3 gene participates in the proliferation of pancreatic cells – decreased expression of the TFF3 gene leads to decreased proliferation of pancreatic -cells, while increased expression leads to increased proliferation of pancreatic -cells, having no influence on their function (44). The TFF3 protein is also related to angiogenesis, which makes it an important factor in tumor pathogenesis (45). Research has identified increased expression of the TFF3 gene in gastrointestinal and lung tumors, advanced prostate cancer, hepatocellular carcinoma and other tumors (46-49). Expression of the TFF3 gene has a predictive role in breast tumors (50) and is simultaneously identified as a valuable and easily detected biomarker in screening for stomach cancers. Moreover, serum TFF3 might predict gastric cancer more efficiently than the PG test, while the combined testing of serum PG (pepsinogen test) and TFF3 could make gastric cancer screening even more efficient (51).

### **Conclusions**

Despite the fact that not much is known about the TFF signaling pathways, some straightforward benefits of TFF peptides for healthy and damaged tissue have been discovered. TFFs are pivotal for mucosal protection and repair of epithelial surfaces, and they also have a role in cancer development and progression. Trefoil factors can be used as prognostic markers for different types of carcinoma. However, their biological effects are still unknown. Considering that there are not many studies on the influence of the TFF peptides on vascular reactivity, it would be interesting to find out more about their role in it.

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## References

1. Blin N. Cytoprotective trefoil peptides abound in new functions. *Cell Mol Life Sci* 2005; 62:2907-9.
2. Thim L. Trefoil peptides: from structure to function. *Cell Mol Life Sci* 1997; 53(11-12): 888-903.
3. Nikolaidis NM<sup>1</sup>, Zimmermann N, King NE, Mishra A, Pope SM, Finkelman FD, Rothenberg ME. Trefoil factor-2 is an allergen-induced gene regulated by Th2 cytokines and STAT6 in the lung. *Am J Respir Cell Mol Biol*.2003 Oct; 29(4): 458-64. Epub 2003 Apr 17.
4. Chen YH<sup>1</sup>, Lu Y, De Plaen IG, Wang LY, Tan XD. Transcription factor NF-kappaB signals antianoinic function of trefoil factor 3 on intestinal epithelial cells. *Biochem. Biophys Res Commun* 2000; 274: 576-582.
5. Graness A, Chwieralski CE, Reinhold D, Thim L, Hoffmann W. Protein kinase C and ERK activation are required for TFF-peptide-stimulated bronchial epithelial cell migration and tumor necrosis factor-alpha-induced interleukin-6 (IL-6) and IL-8 secretion. *J Biol Chem* 2000; 277: 18440-6.
6. Zhu YQ<sup>1</sup>, Tan XD. TFF3 modulates NF-kappaB and a novel negative regulatory molecule of NF- kappa B in intestinal epithelial cells via a mechanism distinct from TNF-[alpha]. *Am J Physiol Cell Physiol* 2005; 289; C1085-93.
7. Kjellev S. The trefoil factor family—small peptides with multiple functionalities. *Cell Mol Life Sci* 2009; 66:1350-69.
8. Thim L, Woldike HF, Nielsen PF, Christensen M, Lynch-Devaney K, Podolsky DK. Characterization of human and rat intestinal trefoil factor produced in yeast. *Biochemistry* 1995; 34(14): 4757-64.
9. Hanby AM, Poulsom R, Singh S, Elia G, Jeffery RE, Wright NA. Spasmolytic polypeptide is a major antral peptide: distribution of the trefoil peptides human spasmolytic polypeptide and pS2 in the stomach. *Gastroenterology* 1993;105: 1110-16.
10. Madsen J, Nielsen O, Tornøe I, Thim L, Holmskov U. Tissue localization of human trefoil factors 1, 2, and 3. *J Histochem Cytochem* 2007; 55(5): 505-13.
11. Gött P, Beck S, Machado JC, Carneiro F, Schmitt H, Blin N. Human trefoil peptides: genomic structure in 21q22.3 and coordinated expression. *EJHG* 1996; 4(6): 308-15.
12. Hoffmann W, Jagla W, Wiede A. Molecular medicine of TFF-peptides: from gut to brain. *Histol Histopathol* 2001, 16: 319-334.
13. Matsuoka Y, Pascall JC, Brown KD. Quantitative analysis reveals differential expression of mucin (MUC2) and intestinal trefoil factor mRNAs along the longitudinal axis of rat intestine. *Biochim Biophys Acta-Gene Struct Expression* 1999; 1489: 336-344.
14. Longman RJ<sup>1</sup>, Douthwaite J, Sylvester PA, Poulsom R, Corfield AP, Thomas MG, Wright NA. Coordinated localisation of mucins and trefoil peptides in the ulcer associated cell lineage and the gastrointestinal mucosa. *Gut* 2000; 47: 792-800.
15. Langer G, Jagla W, Behrens-Baumann W, Walter S, Hoffmann W. Ocular TFF-peptides: new mucus associated secretory products of conjunctival goblet cells. *Adv Exp Med Biol* 2002; 506: 313-316.

16. Kouznetsova I, Laubinger W, Kalbacher H, Kalinski T, Meyer F, Roessner A, Hoffmann W. Biosynthesis of gastrokine-2 in the human gastric mucosa: restricted spatial expression along the antral gland axis and differential interaction with TFF1, TFF2 and mucins. *Cell Physiol Biochem* 2007; 20: 899–908.
17. Xue L, Aihara E, Podolsky DK, Wang TC, Montrose MH. In vivo action of trefoil factor 2 (TFF2) to speed gastric repair is independent of cyclooxygenase. *Gut* 2010; 59:1184–91.
18. Taupin D, Wu DC, Jeon WK, Devaney K, Wang TC, Podolsky DK. The trefoil gene family are coordinately expressed immediate-early genes: EGF receptor- and MAP kinase-dependent interregulation. *J Clin Invest* 1999; 103: R31–38.
19. Xue L, Aihara E, Wang TC, Montrose MH. Trefoil factor 2 requires Na/H exchanger 2 activity to enhance mouse gastric epithelial repair. *J Biol Chem* 2011; 286: 38375–82.
20. Aihara E, Engevik KA, Montrose MH. Trefoil Factor Peptides and Gastrointestinal Function. *Annu Rev Physiol* 2017 Feb 10; 79:357–380. doi: 10.1146/annurev-physiol-021115-105447.
21. Shah AA, Leidinger P, Keller A, Wendschlag A, Backes C, Baus-Loncar M, Meese E, Blin N. The intestinal factor Tff3 and a miRNA network regulate murine caloric metabolism. *RNA Biol* 2011; 8(1): 77–81.
22. Ge H, Gardner J, Wu X, Rulifson I, Wang J, Xiong Y, Ye J, Belouski E, Cao P, Tang J, Lee KJ, Coberly S, Wu X, Gupte J, Miao L, Yang L, Nguyen N, Shan B, Yeh WC, Véniant MM, Li Y, Baribault H. Trefoil factor 3 (TFF3) is regulated by food intake, improves glucose tolerance and induces mucinous metaplasia. *PLoS One* 2015; 10(6): e0126924.
23. Guillén N, Navarro MA, Arnal C, Noone E, Arbonés-Mainar JM, Acín S, Surra JC, Muniesa P, Roche HM, Osada J. Microarray analysis of hepatic gene expression identifies new genes involved in Steatotic liver. *Physiol Genomics* 2009; 37(3): 187–98.
24. Xue Y, Shen L, Cui Y, Zhang H, Chen Q, Cui A, Fang F, Chang Y. Tff3, as a novel peptide, regulates hepatic glucose metabolism. *PLoS One* 2013; 8(9): e75240.
25. Barrera Roa GJ, Tortolero GS, Gonzalez JE. Trefoil factor 3 (TFF3) expression is regulated by insulin and glucose. *Journal of Health Sciences* 2013; 3(1): 1–12.
26. Rui L. Energy metabolism in the liver. *Compr Physiol* 2014; 4: 177–197.
27. Liu J, Han L, Zhu L, Yu Y. Free fatty acids, not triglycerides, are associated with non-alcoholic liver injury progression in high fat diet induced obese rats. *Lipids Health Dis* 2016; 15:27.
28. Wang D, Wei Y, Pagliassotti MJ. Saturated fatty acids promote endoplasmic reticulum stress and liver injury in rats with hepatic steatosis. *Endocrinology* 2006; 147: 943–951.
29. Schug TT, Li X. Sirtuin 1 in lipid metabolism and obesity. *Ann Med* 2011; 43: 198–211.
30. Poulsen L la C, Siersbæk M, Mandrup S. PPARs: Fatty acid sensors controlling metabolism. *Semin Cell Dev Biol* 2012; 23: 631–639.
31. Bujak M, Tartaro Bujak I, Sobočanec S, Mihalj M, Novak S, Čosić A, Tolušić Levak M, Kopačin V, Mihaljević B,

- Balog T, Drenjančević I, Lončar MB. Trefoil Factor 3 Deficiency Affects Liver Lipid Metabolism. *Cellular physiology and biochemistry* 47 (2018), 2; 827-841.
32. Liu SQ, Tefft BJ, Roberts DT, Zhang LQ, Ren Y, Li YC, Huang Y, Zhang D, Phillips HR, Wu YH. Cardioprotective proteins upregulated in the liver in response to experimental myocardial ischemia. *Am J Physiol Heart Circ Physiol* 2012; 303(12): H1446-58.
  33. Liu SQ, Roberts D, Zhang B, Ren Y, Zhang LQ, Wu YH. Trefoil factor 3 as an endocrine neuroprotective factor from the liver in experimental cerebral ischemia/reperfusion injury. *PLoS One* 2013; 8(10):e77732.
  34. Drenjančević I, Ćosić A, Novak S, Stupin A, Jukić I, Baus Lončar M, Mihalj M, Mihaljević Z. The influence of high salt intake on carotid arteries responses to acetylcholine and changes in flow in wild type (WT) 129/SV and TFF3<sup>-/-</sup> mice. (abstract) *Proceedings of The Physiological Society* 2016. Dublin, Ireland.
  35. Mihaljević Z, Matić A, Rašić L, Stupin A, Jukić I, Baus- Lončar M, Drenjančević I. Vascular NO and Superoxide Production is Influenced by High Dietary Salt Intake in TFF3<sup>-/-</sup> Mice. (abstract) *Journal of Hypertension* 2018; 36:e201-e202.
  36. Podolsky DK, Gerken G, Eyking A, and Cario E. Colitis-associated variant of TLR2 causes impaired mucosal repair due to TFF3 deficiency. *Gastroenterology* 2009; 137(1): 209-220.
  37. Paulsen FP, Woon CW, Varoga D, Jansen A, Garreis F, Jäger K, et al. Intestinal trefoil factor/TFF3 promotes re-epithelialization of corneal Wounds. *J Biol Chem* 2008; 283(19): 13418-27.
  38. Kouznetsova I, Chwieralski CE, Bälder R, Hinz M, Braun A, Krug N, Hoffmann W. Induced trefoil factor family 1 expression by trans-differentiating Clara cells in a murine asthma model. *Am J Respir Cell Mol Biol* 2007 Mar; 36(3): 286-95. Epub 2006 Sep 21.
  39. LeSimple P, van Seuning I, Buisine MP, Copin MC, Hinz M, Hoffmann W, Hajj R, Brody SL, Coraux C, Puchelle E. Trefoil factor family 3 peptide promotes human airway epithelial ciliated cell differentiation. *Am J Respir Cell Mol Biol* 2007 Mar;36(3):296-303. Epub 2006 Sep 28.
  40. Samson MH, Vestergaard EM, Milman N, Poulsen SS, Nexø E. Circulating serum trefoil factors increase dramatically during pregnancy. *Scand J Clin Lab Invest* 2008; 68(5): 369-74.
  41. Bijelić N, Belovari T, Baus Lončar M. Trefoil factor family protein 3 (TFF3) is present in cartilage during endochondral ossification in the developing mouse fetus. *Acta Histochem* 2013; 115(3): 204-8.
  42. Bijelić N, Perić Kačarević Ž, Belovari T, Radić R. Trefoil factor family protein 3 affects cancellous bone formation in the secondary centers of ossification of mouse tibiae. *Periodicum biologicum* 2015; 117(1): 59-64.
  43. Lubka M, Müller M, Baus-Lončar M, Hinz M, Blaschke K, Hoffmann W, Pfister M, Löwenheim H, Pusch CM, Knipper M, Blin N. Lack of Tff3 peptide results in hearing impairment and accelerated presbycusis. *Cell Physiol Biochem* 2008; 21(5-6): 437-44.
  44. Fueger PT, Schisler JC, Lu D, Babu DA, Mirmira RG, Newgard CB, Hohmeier HE. Trefoil factor 3 stimulates human and rodent pancreatic islet beta-cell replication

- with retention of function. *Mol Endocrinol* 2008; 22(5): 1251-9.
45. Dhar DK, Wang TC, Tabara H, Tonomoto Y, Maruyama R, Tachibana M, Kubota H, Nagasue N. Expression of trefoil factor family members correlates with patient prognosis and neoangiogenesis. *Clin Cancer Res*. 2005;11(18):6472-8.
46. Im S, Yoo C, Jung JH, Choi HJ, Yoo J, Kang CS. Reduced expression of TFF1 and increased expression of TFF3 in gastric cancer: correlation with clinicopathological parameters and prognosis. *Int J Med Sci*. 2013;10(2):133-40.
47. Qu Y, Yang Y, Ma D, Xiao W. Increased trefoil factor 3 levels in the serum of patients with three major histological subtypes of lung cancer. *Oncol Rep* 2012;27(4):1277- 83.
48. Vestergaard EM, Borre M, Poulsen SS, Nexø E, Tørring N. Plasma levels of trefoil factors are increased in patients with advanced prostate cancer. *Clin Cancer Res* 2006;12: 807-12.
49. Okada H, Kimura MT, Tan D, Fujiwara K, Igarashi J, Makuuchi M, Hui AM, Tsurumaru M, Nagase H. Frequent trefoil factor 3 (TFF3) overexpression and promoter hypomethylation in mouse and human hepatocellular carcinomas. *Int J Oncol* 2005; 26(2): 369-77.
50. May FE, Westley BR. TFF3 is a valuable predictive biomarker of endocrine response in metastatic breast cancer. *Endocr Relat Cancer* 2015; 22(3): 465-79.
51. Huang Z, Zhang X, Lu H, Wu L, Wang D, Zhang Q, Ding H. Serum trefoil factor 3 is a promising non-invasive biomarker for gastric cancer screening: A monocentric cohort study in China. *BMC Gastroenterology* 2014; 14:74.